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# Remarks:

The applicant has subsequently filed a sequence listing and declared, that it includes no new matter.

## (54) Cellulose synthase gene

(57) mRNA was extracted at the stage for cotton plant fibrous cells to accumulate cellulose, and cDNA's complementary thereto were synthesized to construct a cDNA library. Clones of a number of 750 were arbitrarily selected from the library, and they were randomly subjected from to sequencing. Those having homology to

an amino acid sequence deduced from a gene of cellulose 4- $\beta$ -glucosyltransferase (bcsA) of cellulose synthase operon of acetic acid bacterium were selected from obtained nucleotide sequences of the respective clones. Thus, DNA coding for cellulose synthase was obtained.

#### Description

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### Technical Field

The present invention relates to a DNA coding for cellulose synthase originating from cotton plant (Gossypium hirsutum), a recombinant DNA containing the same, a transformed cell transformed with the DNA, and a method for controlling cellular cellulose synthesis.

#### Background Art

Cellulose is used for paper, woody structural materials, fiber, cloths, food, cosmetics, and pharmaceuticals, as well

in biosynthesis of cellulose. The cellulose-related industry has been hitherto directed to such cellulose products that have been already produced, in which there has been no trial to develop a new material based on an aspect of biosynthesis. The mechanism of disease action, which is exerted by pathogenic microorganisms on plants, often results from the inhibition on cellulose biosynthesis as in <u>Pyricularia oryzae</u> (<u>P. oryzae</u>). Therefore, the addition of disease resistance to the cellulose biosynthesis mechanism is agriculturally applicable and valuable. Further, cellulose is the most abundant organic compound on the earth, and it is a sink in which the largest amount of CO<sub>2</sub> in the atmospheric air is fixed. Therefore, the genetic improvement of cellulose biosynthesis enzymes is also applicable to the industry which is directed to the control of CO<sub>2</sub> in the atmospheric air based on the use of cellulose as the sink.

In recent years, cDNA's originating from fiber cells of cotton plant have been randomly sequenced, and it has been reported that full length CelA1 and partial length of CelA2 probably represent cDNAs of cotton plant cellulose synthase, in view of the homology to bacterial cellulose synthase gene (bacterial BcsA) (Pear et al., <u>Proceeding of National Academy of Science, USA</u> (1996) <u>93</u> 12637-12642). The binding ability to UDP-glucose has been demonstrated for CelA1. However, as for CelA2, the homology has been merely demonstrated for the C-terminal amino acid sequence.

### Disclosure of the Invention

The present invention has been made in order to provide a new method for regulating cellulose production in prokaryotic cells or eukaryotic cells, an object of which is to provide a DNA coding for cellulose synthase, a recombinant DNA containing the same, a transformed cell transformed with the DNA, and a method for regulating cellular cellulose synthesis.

The present inventors firstly extracted mRNAs at the stage for cotton plant fiber cells to accumulate cellulose, and cDNAs complementary thereto were synthesized to construct a cDNA library. 750 of cDNA clones were arbitrarily selected from the library, and they were randomly subjected to sequencing. Six amino acid sequences were derived for one nucleotide sequence of each of the obtained clones to select those having homology to an amino acid sequence obtained by translation from a gene of cellulose 4-β-glucosyltransferase (bcsA) of cellulose synthase operon of aceto-bacterium. As a result, genes, which were classified into three types or groups, were found, and they were designated as PcsA1, PcsA2, and PcsA3 respectively (PcsA is an abbreviation of "Plant Cellulose Synthase A").

That is, the present invention lies in a DNA coding for any one of the following proteins (A) to (C):

- (A) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 2 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 2;
- (B) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 4 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 4: and
- (C) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 8 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 8, and comprising an amino acid sequence shown in SEQ ID NO: 11 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 11.

In another aspect, the present invention provides a recombinant vector comprising all or a part of the DNA as defined above, and a transformed cell transformed with the DNA as defined above.

In still another aspect, the present invention provides a method for regulating cellulose synthesis in a cell, comprising the steps of introducing the DNA as defined above into the cell, and expressing RNA having a nucleotide

sequence homologous to the DNA as defined above or a nucleotide sequence complementary to the DNA as define above.

SEQ ID NO: 1 corresponds to a sequence of PcsA1, and SEQ ID NO: 3 corresponds to a sequence of PcsA2. SEQ ID NO: 5 corresponds to a sequence of 3'-side region of PcsA3, SEQ ID NO: 7 corresponds to a sequence of 5'-side region of PcsA3, and SEQ ID NO: 9 corresponds to a sequence of internal region of PcsA3.

It has been demonstrated that PcsA1 and PcsA2 of the DNA's described above are DNA's coding for cotton plant cellulose synthase, according to the expression in eukaryotic cells (animal cells and/or yeast). It has been also demonstrated that an antibody thereagainst inhibits the cotton plant cellulose synthase activity in a cell-free system. Further, PcsA3, which is different from PcsAI and PcsA2, has been found. Any one of these species was obtained as partial one, at the stage of clones obtained by the random sequencing, and no 5'-portion of the coding region was contained. Therefore, clones which have sequences of 5'-portions were isolated in accordance with the 5'-RACE method based on the use of PCR to determine the sequences. As a result of this operation, the sequences of the 5'-portions corresponding to the partial length clones were obtained for PcsA1 and PcsA2.

On the other hand, as for PcsA3, a sequence of a 5'-portion of another clone, which was considered to belong to the same PcsA3 group, was obtained. The both sequences had extremely high homology, and hence they were considered to have underwent multiple gene formation relatively recently originating from an identical gene through the process of duplication. Therefore, even when the both are combined with each other at corresponding portions to construct a fused gene followed by expression, it is assumed that the activity and function of a produced enzyme may not be affected thereby.

As for PcsA1 and PcsA2, in order to obtain a full length clone, primers were designed on the basis of the sequence of the 5'-portion and the sequence of the 3'-portion of the partial length clone to perform PCR. Thus, a clone containing ORF was obtained.

Those applicable as the template to be used for the RACE method may be any of cDNA synthesized from mRNA and a phage library. When the phage library is used, it is possible to use a sequence in the vector as a 5'-side primer.

As a result of random sequencing, seven clones concerning PcsA2 were most abundantly present, of 15 clones seemed to code the cellulose synthase. Expression was confirmed in eukaryotic cells (animal cells and/or yeast) transformed with the cellulose synthase gene. As a result, the cellulose synthase activity was observed.

The present invention will be explained in detail below.

### <1> Preparation of cotton plant cDNA library

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Cotton plant fiber cells at the stage of cellulose accumulation are preferably used as a material for extracting mRNA to construct a cotton plant cDNA library. The method for extracting mRNA is not specifically limited, for which it is possible to adopt an ordinary method for extracting mRNA from plant.

cDNA can be synthesized, for example, by using a poly T sequence which is complementary to poly A nucleotide existing at the terminal of mRNA as a primer to synthesize complementary DNA by the aid of reverse transcriptase, and forming a double strand by the aid of DNA polymerase.

The method therefor is described, for example, in <u>Molecular Cloning</u> (Maniatis et al., Cold Spring Harbour Laboratory). However, a variety of cDNA synthesis kits are commercially available from various companies, which may be used.

Generally, the library is constructed by using a phage vector. A variety of commercially available vectors are usable. However, it is preferable to use a vector, for example,  $\lambda$ ZAP vector in which it is unnecessary to perform recloning from the vector, and it is possible to immediately prepare a plasmid for sequencing.

# <2> Determination of nucleotide sequence of cDNA

Clones are randomly selected from the obtained cDNA library to determine nucleotide sequences of inserts in the clones. The nucleotide sequence can be determined in accordance with the Maxam-Gilbert method or the dideoxy method. Among them, the dideoxy method is more convenient and preferred.

The nucleotide sequence can be determined in accordance with the dideoxy method by using a commercially available sequencing kit. Further, the use of an automatic sequencer makes it possible to determine sequences of a large number of clones for a short period of time.

It is unnecessary to determine the sequence for an entire length of the insert. It is enough to determine a length of nucleotide sequence which is considered to be sufficient to perform homology search. For example, in Examples described later on, the homology search as described below was performed when a sequence having not less than 60 nucleotides was successfully determined.

## <3> Homology search with gene data base

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The determined nucleotide sequence of each of cDNA clones is used to perform the homology search with respect to known amino acid sequences of the cellulose synthase or nucleotide sequences of genes coding therefor registered in the gene data base. The cellulose synthase is exemplified by an enzyme encoded by a gene of cellulose 4β-glucosyltransferase (BcsA) of cellulose synthase operon of acetobacterium (Wong, H. C. et al., Proc. Natl. Acad. Sci. U.S.A., 87, 8130-8134 (1990). ACCESSION No. M37202).

Those usable as the data base include, for example, GenBank, EMBL, and DDBJ published, for example, from Los Alamos National Institute in the United States, Institute of European Molecular Biology, and National Institute of Genetics (Japan). Those commercially available and useable as the program for homology search include, for example, commercially available DNA analysis softwares, such as DNASIS (Hitachi Software Engineering Co.,Ltd.) and GENE-

nected on Internet with NCBI (National Center for Biotechnology Information) to utilize (http://www.ncbi.nim.mn.gov/BLAST/) BLAST (Basic Local Alignment Search Tool) so that high speed homology search is performed.

The homology search is performed, for example, in accordance with the following algorithm. When the homology search is performed for a nucleotide sequence, homology comparison is advanced while shifting the nucleotide sequence to be investigated by every one nucleotide with respect to individual gene sequences included in the data base. When six or more continuous nucleotides are coincident, the homology score is counted and calculated in accordance with a homology score table (see, for example, M. Dayhoff, Atlas of Protein Sequence and Structure, vol. 5 (1978)). The system is set so that those having a score not less than a certain value are picked up as candidates which have homology. Further, the gap may be introduced into the sequence to be investigated or into the gene sequence included in the data base to make optimization so that the score is maximized.

When the homology search is performed for an amino acid sequence, a nucleotide sequence to be investigated is converted into amino acids concerning all six frames including those of a complementary chain. The investigation may be performed in the same manner as performed for the nucleotide. Specifically, it is possible to use blastx of BLAST described above. As for detailed techniques and conditions for the search, reference may be made to <u>DDBJ News Letter</u>, No. 15 (February 1995).

#### <4> Isolation of cDNA clone of cotton plant cellulose synthase

The clone obtained as described above is not necessarily contain the entire nucleotide sequence of the gene. In such a case, the clone is used as a probe to perform screening by means of plaque hybridization. Thus, it is possible to obtain a clone containing a full length gene from the library. A specified method may be carried out with reference to Molecular Cloning, second edition (Maniatis et al., Cold Spring Harbour Laboratory) 12.30 to 12.40.

When obtained cDNA is deficient in 5'-portion, the 5'-portion can be obtained as well by synthesizing primers so that the cDNA sequence may be elongated toward the 5'-terminal, and performing RT-PCR by using mRNA as a template

As demonstrated in Examples described later on, the DNA of the present invention has been obtained as those having homology to the known bacterial cellulose synthase gene. The DNA further codes for an amino acid sequence GlnXXXXXXArgTrp (SEQID NO: 12) which is considered to form a UDP-glucose binding domain, having high homology in the vicinity thereof.

The nucleotide sequences of DNA of the present invention obtained as described above and the amino acid sequences deduced from the nucleotide sequences are shown in SEQ ID NOs: 1 to 10 in Sequence Listing. SEQ ID NOs: 1 and 3 show nucleotide sequences of PcsAl and PcsA2 respectively. SEQ ID NOs: 2 and 4 show amino acid sequences deduced from the nucleotide sequences of PcsA1 and PcsA2 respectively.

SEQ ID NOs: 5 and 6 show a nucleotide sequence of a clone (PcsA3-682) containing 3'-side region of PcsA3 and an amino acid sequence deduced from the nucleotide sequence respectively. SEQ ID NOs: 7 and 8 show a nucleotide sequence of a 5'-portion (PcsA3-5') of another clone containing 5'-side region of PcsA3 and an amino acid sequence deduced from the nucleotide sequence respectively. SEQ ID NOs: 9 and 10 show a nucleotide sequence of 3'-portion (PcsA3-3') of the clone and an amino acid sequence deduced from the nucleotide sequence respectively (see Fig. 1). That is, SEQ ID NO: 5 corresponds to the 3'-side region of PcsA3, SEQ ID NO: 7 corresponds to the 5'-side region of PcsA3, and SEQ ID NO: 9 corresponds to internal region of PcsA3. The overlapping portion of PcsA3-682 is different from that of PcsA3-3' in 9 nucleotides in the nucleotide sequence and 1 amino acid in the amino acid sequence. Figs. 3 and 4 show the comparison between the nucleotide sequences of PcsA3-682 and PcsA3-3'. SEQ ID NO: 11 shows a combination of the amino acid sequences encoded by PcsA3-682 and PcsA3-3'.

The sequence of GInXXXXXXArgTrp (SEQ ID NO: 12) corresponds to amino acid numbers 710 to 714 in SEQ ID NO: 2 for PcsA1, amino acid numbers 778 to 782 in SEQ ID NO: 4 for PcsA2, and amino acid numbers 356 to 360 in

	530 535 540
	His Asp Arg Tyr Ala Asn Arg Asn Val Val Phe Phe Asp Ile Asn Met
5	
	Leu Gly Leu Asp Gly Leu Gln Gly Pro Val Tyr Val Gly Thr Gly Cys  565  570
10	Val Phe Asn Arg Gln Ala Leu Tyr Gly Tyr Asp Pro Pro Val Ser Glu
10	785
	bys Arg Pro Lys Met Thr Cys Asp Cys Trp Pro Ser Trp Cys Cys Cys
	600
	Cys Cys Gly Gly Ser Arg Lys Lys Ser Lys Lys Lys Cly Cly Lee
15	615
	Gly Leu Leu Gly Gly Leu Leu Tyr Gly Lys Lys Lys Met Met Gly
	030
	Lys Asn Tyr Val Lys Lys Gly Ser Ala Pro Val Phe Asp Leu Glu Glu
20	645 650
•	
	Ile Glu Glu Gly Leu Glu Gly Tyr Glu Glu Leu Glu Lys Ser Thr Leu  660
	UDD 670
25	Met Ser Gln Lys Asn Phe Glu Lys Arg Phe Gly Gln Ser Pro Val Phe
	680
	Ile Ala Ser Thr Leu Met Glu Asn Gly Gly Leu Pro Glu Gly Thr Asn
	095 700
30	Ser Thr Ser Leu Ile Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr
	710 715
	Glu Glu Lys Thr Glu Trp Gly Lys Glu Ile Gly Trp Ile Tyr Gly Ser
	725 730
35	Val Thr Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys Arg Gly Trp
	745
	Lys Ser Val Tyr Cys Val Pro Lys Arg Pro Ala Phe Lys Gly Ser Ala
40	Pro Ile Asn Leu Ser Asp Arg Leu His Gln Val Leu Arg Trp Ala Leu 770
	Gly Ser Val Glu Tle Pho Tou San 3
	Gly Ser Val Glu Ile Phe Leu Ser Arg His Cys Pro Leu Trp Tyr Gly
45	705
	Tyr Gly Gly Lys Leu Lys Trp Leu Glu Arg Leu Ala Tyr Ile Asn Thr
	900
	The Fire The Ser Ile Pro Leu Ala Tyr Cyc The Ile
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	Asn Leu Thr Ser Val Trp Phe Leu Ala Leu Phe Leu Ser Ile Ile Ala
<i>55</i>	
	860

	Thr Gly Val Leu Glu Leu Arg Trp Ser Gly Val Ser Ile Gln Asp Trp 875 880	
	865	
5	Trp Arg Asn Glu Gln Phe Trp Val Ile Gly Gly Val Ser Ala His Leu	
•	885	
	Phe Ala Val Phe Gln Gly Leu Leu Lys Val Leu Ala Gly Val Asp Thr	
	900 905 910	
	Asn Phe Thr Val Thr Ala Lys Ala Ala Asp Asp Thr Glu Phe Gly Glu	
10	915 920 925	
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_	Ile Ile Leu Asn Met Val Gly Val Val Ala Gly Val Ser Asp Ala Ile	
15	945 950 955 960	
	Asn Asn Gly Tyr Gly Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe	
	965 970 975	
	Ala Phe Trp Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly Leu Met	
20	980 985 990	
	Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Val Leu Trp Ser Ile Leu	
	995 1000 1005	
	Leu Ala Ser Ile Phe Ser Leu Val Trp Val Arg Ile Asp Pro Phe Leu	
25	1010 1015 1020	
	Pro Lys Gln Thr Gly Pro Val Leu Lys Gln Cys Gly Val Glu Cys	
	1025	
	1025	
30	(2) INFORMATION FOR SEQ ID NO: 5:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 2033 base pairs	
	(B) TYPE: nucleic acid	
35	(C) STRANDEDNESS: double	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: CDNA to mRNA	
	(vi) ORIGINAL SOURCE:	
40	(A) ORGANISM: Gossypium hirsutum L.	
	(C) INDIVIDUAL ISOLATE: Coker312	
	(ix) FEATURE: (A) NAME/KEY: CDS	
45	·	
	(B) LOCATION: 1 1857	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:	
	COG ACA TTC GTG AAG GAG GGT GGA GCT ATG AAG AGA GAA THE GET GET	
50	Pro Thr Phe Val Lys Glu Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu	
50	1	
	TTC AAG GTT AGG ATA AAT GCA CTT GTA GOC AAA GOC CAA AAG GTT OCT 96	

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			35					40	)				4	5			
	ACT	AAA	GAT	CAC	CCT	GGT	ATG	ATT	CAA	GTA	TIT	CIC	CCT	CAA	AGT	GGA	192
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	Gly	His	Asp	Thr	Glu	Gly	Asn	Glu	Leu	Pro	Arg	Leu	Val	Tyr	Val	Ser	
15	65					70					7	-				80	
	CCA	GAG	AAA	AGG	CCT	CCT	TTC	TTG	CAT	CAC	AAG	AAA	CCT	CCT	$\infty$	ATG	288
	Arg	Glu	Lys	Arg	Pro	Gly	Phe	Leu	His	His	Lys	Lys	Ala	Gly	Ala	Met	
					85					90	-					5	
20					$\infty$												336
	Asn	Ala	Leu	Val	Arg	Val	Ser	Gly	Val	Leu	Thr	Asn	Ala	Pro	Phe	Met	
				100					105					11			
					TGT												384
25	Leu	Asn	Leu	yab	Cys	Asp	His	Tyr	Leu	Asn	Asn	Ser			Val	Arg	
			115					120					12				
					TTC												432
	Glu	Ala	Met	Cys	Phe	Leu	Met	Asp	Pro	Gln	Ile	Gly	Arg	Lys	Val	CAa	
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	Tyr	Val	Gln	Phe	Pro	Gln	Arg	Phe	Asp	Gly		_	Arg	His	Asp		
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35					AAC												528
	Tyr	Ala	Asn	Arg	Asn		Val	Phe	Phe			Asn	Met	Lys			
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					GGC												576
40	Asp	Gly	Ile		Gly	Pro	Val	Tyr			Thr	Gly	Cys	_	_	Arg	
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					TAT												624
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	Lys		Val	Thr	Cys	Gly			Pro	Cys	Phe			Arg	Arg	rys	
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	Asp	Lys	Lys	His	Ser		Asp	Gly	Gly	Asn			GLY	Leu	Ser		
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	GAA	GCA	œ	AAA	GAT	GAC	AAG	GAG	TTA	TTG	ATG	TOO	CAC	ATG	AAC	TTT	768
	Glu	Ala	Ala	Lys	Asp	Asp	Lys	Glu	Leu	Leu	Met	Ser	His	Met	Asn	Phe	
5					245	5				25	0				25	55	
	GAA	AAG	AAA	TTT	GGA	CAA	TCA	GCC	ATT	TTT	GTA	ACT	TCA	ACA	CIG	ATG	816
	Glu	Lys	Lys	Phe	Gly	Gln	Ser	Ala	Ile	Phe	Val	Thr	Ser	Thr	Leu	Met	
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					Cys												
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	ACA	TCT	TTA	CCA	CTT	CTC	ccc	TAT	TGT	ACC	CTA	$\infty$	GCA	ATC	TGT	TTA	1248
40	Thr	Ser	Leu	Pro	Leu	Leu	Ala	Tyr	Cys	Thr	Leu	Pro	Ala	Ile	Cys	Leu	
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	CIT	ACC	GAT	AAA	TTT	ATC	ATG	CCA	$\infty$	ATA	AGC	ACC	TTT	GCA	AGT	CTA	1296
	Leu	Thr	Asp	Lys	Phe	Ile	Met	Pro	Pro	Ile	Ser	Thr	Phe	Ala	Ser	Leu	
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40	TTC	TTC	ATT	$\infty$	TTG	TTT	CIT	TCA	ATC	TTT	GCA	ACT	CCT	ATT	CTC	GAG	1344
	Phe	Phe	Ile	Ala	Leu	Phe	Leu	Ser	Ile	Phe	Ala	Thr	Gly	Ile	Leu	Glu	
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5 <b>0</b>	CTA																1392
50	Leu	Arg	Trp	Ser	Gly	Val	Ser	Ile	Glu	Glu	Trp	Trp	Arg	Asn	Glu	Gln	

		450					455	i				460	)				
	TTT	TGG	GTC	ATC	GGT	GCC	ATT	TCG	GCA	CAT	TTG	TTC	CCT	GTT	ATC	CAA	1440
	Phe	Trp	Val	Ile	Gly	Gly	Ile	Ser	Ala	His	Leu	Phe	Ala	Val	Ile	Gln	
5	465					470	<b>,</b>				475	5				480	
	œc	TTG	TTG	AAA	GTT	CTA	CCI	GGT	ATT	GAC	ACT	AAT	TTC	ACT	GIC	ACA	1488
	Gly	Leu	Leu	Lys	Val	Leu	Ala	Gly	Ile	Asp	Thr	Asn	Phe	Thr	Val	Thr	
10					485	i				490	)				49	5	
10	TCC	AAG	GCA	ACT	GAT	GAC	GAG	GAG	TTC	GGG	GAA	TTG	TAT	ACT	TIC	AAA	1536
	Ser	Lys	Ala	Thr	Asp	Asp	Glu	Glu	Phe	Gly	Glu	Leu	Tyr	Thr	Phe	Lys	
				500					505					51	_		
			ACC														1584
15	Trp	Thr	Thr	Leu	Leu	Ile	Pro	Pro	Thr	Thr	Val	Leu	Ile	Ile	Asn	Leu	
			515					520					52				
			GIC														1632
	Val	Gly	Val	Val	Ala	Gly	Ile	Ser	Asp	Ala	Ile	Asn	Asn	Gly	Tyr	Gln	
20		530					535					540					
			GGA														1680
	Ser	Trp	Gly	Pro	Leu	Phe	Gly	Lys	Leu	Phe	Phe	Ser	Phe	Trp	Val		
	545					550					555					560	
25																	1728
	Val	His	Leu	Tyr	Pro	Phe	Leu	Lys	Gly			Gly	Arg	Gln		_	
			CTC TAT CCA T Leu Tyr Pro F 565 ACC ATT GTT C Thr Ile Val V							570					57		
																	1776
30	Thr	Pro	Thr			Val	Ile	Trp			Leu	Leu	Ala			Phe	
				580					585					590	-		1004
			CIT														1824
	Ser	Leu	Leu	Trp	Val	Arg	Ile			Phe	Val	Met			Lys	GTĀ	
35			595			_		600					60				1004
			ACT	-							TGA	\AAAA,	AAA 1	CATC	ZI'IGC	X3	1874
	Pro		Thr	Thr	Met	Cys			Asn	Cys							
		610					615		max				~~~a	<b>~</b>	~~~	2000	1024
40																AAGAC	1934 1994
											_	CATT	CIAL	CA A	CTAT	AAGIT	
	TIGI	CATI	CA A	VI TGA	LAAA'I	'A GC	JICAA	CTT	GIG	ATCA	AA						2033
	(2)	TNEY	emai	יאחאי	EVP	SEX	א חז	n· e									
45	(2)		SEC														
		( 1		-		#: 6:				łe							
			-	•		ami			سد								
			-			OGY:											
50		(44)	IOM	•													
		( )					الوسي										

(v) FRAGMENT TYPE: C-terminal fragment

		(x1)	) SEX	UEN	Œ DE	SCRI	PTIC	<b>W:</b> S	EQ I	D NC	: 6:						
	Pro	Thr	Phe	Val	Lys	Glu	Arg	Arg	Ala	Met	Lys	Arg	Glu	Tyr	Glu	Glu	
5	1				5					10					1		
	Phe	Lys	Val	Arg	Ile	Asn	Ala	Leu	Val	Ala	Lys	Ala	Gln	_	_	Pro	
				20					25					3		_	
10	Pro	Glu	_	Trp	Ile	Met	Gln			Thr	Pro	Trp			Asn	Asn	
		uniteral Park	35			a series and	-	40			duranti lovo la	mbuti Demilia	45				
		50					55					60					
	Gl <sub>17</sub>	His	Acro	Thr	Glu	Glv	-		ī <sub>en</sub>	Pm	Am			Tvr	Val	Ser	
15	65	urs	nap	1111	Giu	70		014		110	75	_	•—	-4-		80	
		Glu	Lus	Am	Pro			Leu	His	His			Ala	Gly	Ala	Met	
	9	020	-10	9	85					90		-		_	9	,	
20	Asn	Ala	Leu	Val			Ser	Gly	Val	Leu	Thr	Asn	Ala	Pro	Phe	Met	
				100				_	105					110			
	Leu	Asn	Leu	Asp	Cys	Asp	His	Tyr	Leu	Asn	Asn	Ser	Lys	Ala	Val	Arg	
			115					120					129	5			
<i>25</i> .	Glu	Ala	Met	Cys	Phe	Leu	Met	Asp	Pro	Gln	Ile	Gly	Arg	Lys	Val	Cys	
		130					135	;				140	)				
	Tyr	Val	Gln	Phe	Pro	Gln	Arg	Phe	Asp	Gly			Arg	His	Asp		
30	145					150					155					160	
	Tyr	Ala	Asn	Arg			Val	Phe	Phe			Asn	Met	Lys			
	_				165		17-1		**-1	170		C1	<b>ر</b> يد	บอไ	17 Pho	_	
	ASP	Gly	TTE	180		PIO	vai	TYL	185		1111	GTĀ	Cys	19		nig .	
35	<b>3</b>	Gln	<b>8</b> 1.0			C117	(Terror	Glu.			Lare	Glv	Pm			Pm	
	Arg	GIII	195		TĂT	GLY	ıyı	200		110	Ly S	CLY	209		9		
	Lve	Met			Cvs	Glv	Cvs			Cvs	Phe	Gly			Arg	Lys	
40	,-	210			-1-		215			•		220			_	_	
	Asp	Lys	Lys	His	Ser	Lys	Asp	Gly	Gly	Asn	Ala	Asn	Gly	Leu	Ser	Leu	
	225					230	)				235	5				240	
	Glu	Ala	Ala	Lys	Asp	Asp	Lys	Glu	Leu	Leu	Met	Ser	His	Met	Asn	Phe	
45					245	i				250	)				25	5	
	Glu	Lys	Lys	Phe	Gly	Gln	Ser	Ala	Ile	Phe	Val	Thr	Ser			Met	
				260					265					27		_	
50	Glu	Gln	Gly	Gly	Val	Pro	Pro			Ser	Pro	Ala			Leu	rys	
			275				_	280				_	28		<b>~</b> 3	<b>—</b>	
	Glu	Ala		His	Val	Ile			Gly	Tyr	GLu			TUT	GIU	dı	
		290		_		<b></b>	295		<b>6</b> 7	<b>.</b>	T1 -	300		<b>&gt;</b>	<b>7</b> 7.	Tou	
55	Gly	Ser	Glu	Leu	GLY	JIP	ΤŢ	JAL	GTA	ser	TTE	INT	GIU	ASD	тте	Leu	

	303					31					3:	15				32	20
	Thr	Gl	y Pha	e Lys	s Met	t His	з Су	s Ar	g Gl	у Тт	) Ar	y Se	r Ile	e Tyr	c Cy	s Met	_
5					32	5				33				4		335	
	Pro	Lys	Let	ı Pro	Ala	. Phe	Ly	s Gl	y Se	r Ala	Pro	o Ile	Ast	Leu	ıSe	r Asp	
				340	)				34						50	r .mp	
	Arg	Leu	ı Ast	ı Glr	Val	Leu	Arg	y Try	) Ala	a Leu	Gly	7 Sex	. Val	Glu	. T]4	e Phe	
10			355	5				36	0		-4		36				
	Phe	Ser	His	His	Cys	Pro	Ala	Tr	Tyr	Glv	Phe	Lvs	្រ ព្រះ	្រ ព្រះ	Tare	s Leu	
		370	).				37	5	-			38		OLY	₽J.	> Leu	
	Lys	Trp	Leu	Glu	Arg	Phe	Ala	Тут	· Val	Asn	Thr	Thr	Tle	Тал	Dr	Phe	
15	385					390	)	-			39				110	40	_
	Thr	Ser	Leu	Pro	Leu	Leu	Ala	Tyr	Cvs	Thr	Leu	Pm	Δla	Tla	C+200	Leu	ر
					405	i		-	- 2	410			rua	116		15	
22	Leu	Thr	Asp	Lys	Phe	Ile	Met	Pro	Pro	Ile	Ser	<b>ጥ</b> ንድ	Pho	Δla	Sor	Leu	
20				420					42					43		Leu	
	Phe	Phe	Ile	Ala	Leu	Phe	Leu	Ser			Ala	Thr	Glv	Jle alī	Ten.	Glu	
			435				~	44(	)				44		Leu	Giu	
25	Leu	Arg	Trp	Ser	Gly	Val	Ser	Ile	Glu	Glu	Trr	רגים	Δτοτ	) Aen	Gl.,	Gln	
		<b>4</b> 50					455	,				460		ASLI	GIU	GIII	
	Phe '	Trp	Val	Ile	Gly	Gly	Ile	Ser	Ala	His	Leu	Phe	Ala	Val	Tla	Gln	
	465					470					475			<b>,</b>	116	480	ı
30	Gly 1	Leu	Leu	Lys	Val	Leu	Ala	Gly	Ile	Asp	Thr	Asn	Phe	Thr	Val	ωρ. ≖οο	
					485					490					40	5	
	Ser I	Lys .	Ala	Thr .	Asp .	Asp	Glu	Glu	Phe	Gly	Glu	Leu	Tvr	Thr	Phe	Lve	
				500					505				-1-	510		273	
35	Trp 1	thr '	Thr	Leu :	Leu :	Ile :	Pro	Pro	Thr	Thr	Val	Leu	Ile	Tle	Asn	Ī 🕒 I I	
			212					520					525				
	Val G	ily v	Val '	Val 2	Ala (	Gly :	Ile	Ser	Asp	Ala :	ile .	Asn .	Asn (	Glv '	ľvr	Gln	
10	=	30		-			535					540					
40	Ser T	dı,	Gly 1	Pro I	Leu I	Phe (	Gly	Lys	Leu :	Phe 1	Phe s	Ser 1	Phe 1	7 ond	<i>l</i> al	Tle	
	343					550					55 <b>5</b>					560	
	Val H	is I	eu 1	làr i	ro F	he I	eu i	Lys (	Gly i	Leu N	æt (	ily A	ara (	3ln A	lsn i	Am	
45				,	565					570					575	:	
	Thr P	ro T	hr 1	le V	al V	al I	le 1	Crp s	Ser 1	Val I	eu I	eu A	la S	er I	le I	Phe	
				580					585					590		••~	
	Ser Le	eu L	eu 1	λ đτ	al A	rg I	le A	Asp I	oro	he v	al M	let I	vs 1	hr L	vs (	210	
50		5	95					600		•			605		, 0	I	
	Pro As	sp Ti	hr T	hr M	et C	ys G	ly I	le A	sn C	ys							
	61						515			-							
	<b></b>																
55	(2) IN	IFOR	ITAN	ON FO	OR SI	EQ II	OM C	: 7:									

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1086 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: double(D) TOPOLOGY: linear

		(11	) MOI	ECUI	LE T	PE:	CDN	A to	mRN	A							
10		(vi	) OR	(GIN	AL S	OURCE	Ξ:										
			(2	A) O	RGAN	ISM:	Cos	sypi	m h	irsut	tum 1	<b>L.</b>					
		(ix	) FE	TURE	<b>:</b> :							,,,					
15			(.	A) N	AME/	KEY:	$\cos$										
			•	B) L													
			) SE(														
	GGC	ACGA(	CT 1	MCA!	CATC	$x \propto$											50
20							۲	et G	lu A	ıla S	ier A	la C	ly I	eu V	al A	la	
								1				5					
			CAC														98
	Gly	Ser	His	Asn	Arg	Asn	Glu	Leu	Val	Val			Gly	His	Glu		_
25	10					15					20					25	
			$\infty$ r														1 <b>4</b> 6
	Pro	Lys	Pro	Leu			Leu	Asp	Gly			Cys	Glu	He			
		30 35 40 AT GAA ATT GGG TTG ACG GTC GAT GGA GAT CTT TTC GTG GCC TGC AAC														104	
30																	194
	Asp	Glu	Ile			Thr	Val	Asp		_	Leu	Pne	vaı			ASH	
				45					50		m s m	CNC	mam	_	5 ~~~	303	242
			GGT														242
35	Glu	Cys	Gly		Pro	vaı	Cys			Cys	TYL	GIU	79E	-	ALY	Arg	•
	<i>a</i>	~~~	60 AGT		C3.3	m~m	~~	65 C22		222	۸∕₹۳	aca.	-	-	Can	CIC	290
			Ser	-													250
	GIU	75		GIII	GIII	Cys	80		Cys	пуs	1111	ALG 8		цуз	AL 9		
40	AAC		AGT	œ	ACC	GIK			САТ	GAA	САТ		_	GAT	GIG	GAT	338
			Ser														
	90 90	GIY	Jest	110	· <del>u</del> 9	95		017	·Lp	0_0	100					105	<b>;</b>
		ΔTY	GAA	САТ	GAA			ATT	GAT	GAT		-	AAC	AAG	TAT	AGA	386
45			Glu														
				•—–	110				•	115	_			-	12	_	
	ТАА	ATC	GCT	GAA			CTT	CAT	GGA	AAG	ATG	AGC	TAC	GGG	AGA	GGC	434
			Ala														
50				125					130				-	13		_	
	CCT	GAA	GAC			CCT	TTG	CAA	ATC	CCA	$\infty$	GGT	TTA	CCT	<b>GGT</b>	GTT	482

	Pro	Glu	Asp	Asp	Glu	Gly	Leu	Gln	Ile	Pro	Pro	Gly	Leu	Ala	Gly	Val	
			140					145					15	-			
5						AGC											530
	Arg	Ser	Arg	Pro	Val	Ser	Gly	Glu	Phe	Pro	Ile	Gly	Ser.	Ser	Leu	Ala	
		155					160					16					
						TCA											578
10	Tyr	Gly	Glu	His	Met	Ser	Asn	Lys	Arg	Val			Tyr	Pro	Met		
, 0	170					175					180					185	
						AGA											626
	Glu	Pro	Gly	Ser	Ala	Arg	Trp	Asp	Glu			Glu	Gly	Gly			
15					190					195					20		<b>6</b> 71
.5						TGG											674
	Glu	Arg	Met			Trp	Lys	Met			Gly	Asn	Leu		_	GIU	
				205					210					21			700
20						GAT											722
20	Pro	Asp		Ala	Tyr	Asp	ATS			ALA	met	Leu		_	AIa	Arg	
			220					225		~~		.~~	23		3 3 (0)	~~	770
						AAA											770
25	Gln		Leu	Ser	Arg	Lys			rre	ATa	ser			TTE	ASI	PIO	
25		235				~~~	240		~ma	~~~~	<b>.</b>	24		mana.	(INTER)	Catali	818
						GTG											010
	_	Arg	met	vaı	TTE	Val		Arg	Leu	var	260		wra	FIE	Fire	265	
20	250	mam.	~~	3.000	mme	255 AAC		CTD3	Cam	CATE			ccc	باعلت	TYCE		866
30																	000
	AIG	Tyr	ALG	TTE	270	Asn	PLU	AGT	ms	ASP 275		116	Gry	LEU	28		
	1.CT	my m	conc.	» mc		GAA	אייני מ	WCC.	بالعليك			מיית	TYCYC:	ΔΤΥ			914
25						Glu											744
35	1111	SEL	Val	285	Cys	GIU	116	ııp	290		112	<i>-</i>	11p	29		р	
	CAC	- Alath	COTT.	•	W.	TTC	رس.	ידינים			GAG	ACCG"	ТАТ		_	CCC	962
						Phe											
10	0111		300	<b></b>				305		9			310	_	•		
40	بلبان	TCC		AGG	ТАТ	GAG	AGG			GAG	$\infty$	AAC			GCT	TCT	1010
						Glu											
		315		9	-3-		320					325					
45	GIT		ATT	TTT	GTC	AGT	ACA	GTG	GAT	CCA	TTG	AAG	GGA	CCT	œ	CTA	1058
45						Ser											
	330					335			-		340		-			345	
		ACA	GCG	TAA	ACA	GIT		TCG	ATC	T							1086
50						Val											
50					350												

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

5			(	(A) I	ENG	TH: 3	35 <b>4</b> a	mino	aci	.ds							
			(	(B) 1	YPE:	emi.	no a	cid									
			(	(D) 1	OPOI	OGY:	lin	ear									
		(11	L) MC	LECU	LE T	YPE:	pep	tide									
10		(v)	FRA	GMEN	T TY	PE:	N-te	rmin	al f	ragm	ent						
	and the second s	_(.x:	LSE	) O LIBA	CE_D	ESCR.	I PIL	ON	SEO_	ID_N	02	•					
						-								-			
15	1				5					1						1.5	
	Leu	Val	. Val	Ile		Gly	His	Glu			Lys	Pro	Leu	Lys	Asn	Leu	
				20					2						80		
	Asp	Gly		Val	Cys	Glu	Ile	Cys	Gly	Asp	Glu	Ile	Gly	Leu	Thr	Val	
20			35					40	_				4				
	Asp			Leu	Phe	Val			Asn	Glu	Cys			Pro	Val	Cys	
	_	50		_			55					6					
			Cys	Tyr	Glu			Arg	Arg	Glu			Gln	Gln	Cys		
25	65		<b>-</b>		_	70		_	_	_	7!	_				80	
	GIN	Cys	rys	Thr			Lys	Arg	Leu			Ser	Pro	Arg			
	<b>03</b>	•	<b>61</b>		85		_		_	90						5	
30	GIÀ	Asp	GIU	Asp		GIU	ASp	vaı			TTE	Glu	His			Asn	
	77.	<b>3</b>	<b>3</b>	100		<b>&gt;</b>	•	<b>m</b>	105		-1.			110		_	
	116	ASD	ASP 115	Glu	GIN	ASn	rĀ2	12C		ASI	тте	Ата			Met	Leu	
	Wie.	Gly			Ser	Trans.	C1			Desc	C1	<b>&gt;</b>	125		<b>C</b> 1	T	
35	1113	130		Met	Ser	TYL	135		GIY	PLO	Gru	ASD 14(	_	GIU	GTĀ	Leu	
	Gln			Pro	Glv	LOU			V-1	λ	Sor			v-1	Cor	C1	
	145		110	110	O <sub>T</sub>	150		GIY	var	щg	155		PLO	vaT	Ser	160	
		Phe	Pro	Ile	Glv			Teu	Ala	ጥህጉ			Hic	Mat	Sor		
40					165					170		014			17		
	Lys	Arg	Val	His			Pro	Met	Ser			Glv	Ser	Ala			
	-			180		-4 -			185			<b>0-</b> 2	-	190	_		
45	Asp	Glu	Lys	Lys	Glu	Gly	Gly	Trp			Ara	Met	Asp			Lvs	
	-		195	_		-	•	200					205		<b>F</b>	_1 -	
	Met	Gln	Gln	Gly	Asn	Leu	Gly	Pro	Glu	Pro	Asp	Asp			Asp	Ala	
		210		_			215				•	220		-4-			
50	Asp	Met	Ala	Met	Leu	Asp		Ala	Arg	Gln	Pro			Arg	Lys	Val	
	225					230			_		235			3	-	240	
	Pro	Ile	Ala	Ser	Ser	Lys	Ile	Asn	Pro	Tyr	Arg	Met	Val	Ile	Val	Ala	
					245					250					255		

	Arg	Leu	Val	260		Ala	Phe	Phe			Тух	Arg	Ile			n Pr	က
	۷a۱	Hie	Aen			C1**	Lou	Т	26	-	<b>-</b>				270		
5			<b>Asp</b> 275					280	)				28	35			
	Ттр	Phe 290	Ala	Phe	Ser	Trp	Ile 295		Asp	Gln	Phe	Pro 30		Tr	p Ph	e Pr	.o.
10	Ile 305	Asp	Arg	Glu	Thr	Tyr 310		Asp	Arg	Leu		Leu		Ту	r Gl		_
			Glu	D~~	A ~~			11-	0	· · - 3	319						320
	Ozu	OLY	Glu	PIO	325	MEC	Leu	Ата	ser			Ile	Phe	· Va			<b>r</b>
	Vel	Aen	Pro	Lon		C1	D	D	•	330			_			35	
15	Val	rap	Pro	340	гÃЗ	GIĀ	PIO	PTO			Thr	Ala	Asn	_		l Le	u
	Ser	Ile		340					345	)				3	50		
	(2)	TATES															
20	(2)		RMAT														
٠		(1)	SEQ														
				) LE					_	S							
				) TY ) ST													
25				) TO					те								
		(11)	MOL						-DAD								
			ORI					ω,	HUCOVA								
		( -,		) OR				um i i r	m hi	***	T						
30				) IN								•					
		(ix)	FEA?			-0.11	100	G-11 L	٠ س	vera	12						
				NAN (		Y: (	DS.										
				LOC				00									
35		(xi)	SEQU						20 II	NO:	9:						
	GAC /	AAA C	erc c	og c	OG A	CA T	TC G	TG A	AG (	AG (	CT (	CGA (	CT.	ATG	AAG	AGA	48
	Asp I	Lys V	/al A	rg P	ro T	hr P	he V	al I	ys (	lu A	Arg A	Arg 2	la i	Met	Lvs	Arg	•
	1				5					10	•	J			1		
40	GAA 1	TAT G	¥AA G	AA T	TC A	AG G	TT A	GG A	TA A	AT C	CA C	TT (	TA (	GCC			96
	Glu 1	JAT G	lu G	lu P	he Ly	ys V	al A	rg I	le A	sn A	la I	eu (	/al /	Ala	Lvs	Ala	,
				20				_	25					3			
	CAA A	VAG G	TT O	ct a	CA G	AA G	GG T	GG A	TC A	TG C	'AA G	AT C	egg i			TGG	144
45	Gln L	ys V	al P	ro P	ro Gi	lu G	ly T	rp I	le M	et G	ln A	sp G	Sly 1	Thr	Pro	Tro	
			35					40					45				
	CCA G	GA A	AC A	AT AC	T A	VA G	AT C	AC O	CT G	GT A	TG A	TT C	AA C	TA	TTT	CTC	192
	Pro G	ly A	sn As	sn Th	ur Ly	rs As	sp H	is P	ro G	ly M	et I	le G	ln V	/al	Phe	Leu	_,_
50		50					55					60			-		

	GGT	CAA	AGT	GGA	GCC	CAT	GAT	ACC	GAA	GGA	AAT	GAG	CTT	CI	्र द्वा	CIC	240	
	Gly	Gln	Ser	Gly	Gly	His	Asp	Thr	Glu	Gly	Asn	Glu	Leu	Pro	Arg	Leu		
5	. 65					70	)				7	5				80		
	GIC	TAT	GTA	TCT	CCA	GAG	AAA	AGG	CCA	GGT	TTC	TTG	CAT	CAC	AAG	AAA	288	
	Val	Tyr	Val	Ser	Arg	Glu	Lys	Arg	Pro	Gly	Phe	Leu	His	His	Lys	Lys		
					85	5				9	0				9	95		
10	CCT	GGT	$\infty$	ATG	AAC	$\infty$	CIT	GIT	CGT	GIC	TOG	GGG	GTG	CIT	ACA	AAT	336	
,,	Ala	Gly	Ala	Met	Asn	Ala	Leu	Val	Arg	Val	Ser	Gly	Val	Leu	Thr	Asm		
				- ^ ^														
		CON	TTT	1620	TTO	FIRE	TIC	CHI	TOT	-6-16	CAC	TAT	TIA	AAT	AAC	AGC	384	
15	Ala	Pro	Phe	Met	Leu	Asn	Leu	Asp	Cys	Asp	His	Tyr	Leu	Asn	Asn	Ser		
13			115	;				120	)				12	5				
	AAG	CT	GTA	AGA	GAG	CCT	ATG	TGT	TTC	TTG	ATG	GAC	ccr	CAA	ATT	GGA	432	
	Lys	Ala	Val	Arg	Glu	Ala	Met	Cys	Phe	Leu	Met	Asp	Pro	Gln	Ile	Gly		
		130					135					14	_					•
20	AGA	AAG	GIT	TGC	TAT	CIC	CAA	TTC	CT	CAA	CCT	TTC	GAT	GGT	ATT	GAT	480	
•	_	Lys	Val	Cys	Tyr	Val	Gln	Phe	Pro	Gln	Arg	Phe	Asp	Gly	Ile	Asp		
	145					150					15	_				160		
	AGA	CAT	GAT	CGA	TAT	$\infty$	AAT	œ	AAC	ACA	GIT	TTC	TTT	GAT	ATT	AAC	528	
25	Arg	His	yab	Arg	Tyr	Ala	Asn	Arg	Asn	Thr	Val	Phe	Phe	Asp	Ile	Asn		
					165					170					17			
	ATG	AAA	CCT	CTA	GAT	CCT	ATA	CAA	GGC	CCT	GTA	TAT	GIC	œc	ACG	GGG	576	
	Met	Lys	Gly		Asp	Gly	Ile	Gln	Gly	Pro	Val	Tyr	Val	Gly	Thr	Gly		
30				180					185					19	_			
	TGT	GIT	TTC	AGA	AGG	CAA	GCT	CIT	TAT	GGT	TAT	GAA	CCT	CCA	AAG	GGA	62 <b>4</b>	
	Cys	Val			Arg	Gln	Ala	Leu	Tyr	Gly	Tyr	Glu	Pro	Pro	Lys	Gly		
			195					200					20					
35					AAA												672	
	Pro		Arg	Pro	Lys	Met			CAa	Gly	Cys	Cys	Pro	Cys	Phe	Gly		
		210					215					220						
					GAC												720	
40	_	Arg	Arg	Lys	Asp	_	_	His	Ser	Lys	-	_	Gly	Asn	Ala			
	225					230					235					240		
					GAA												768	
	Gly	Leu	Ser	Leu	Glu		Ala	Glu	Asp			Glu	Leu	Leu				
45					245					250					25			
					GAA												816	
	His	Met	Asn		Glu	Lys	Lys	Phe			Ser	Ala	Ile			Thr		
				260					265					276				
50					GAA												86 <b>4</b>	
50	COT	<b>ፕትጉ</b>	Tan	Mat	Glu	ദിന	Glaz	Clv	いっし	Drr	Dvv	SOT	Sor	Cor	D~	Ala		

			2	75				2	80					285				
	GC.	ייני יוני	rg C	IC AZ	NA G	NA GC	X AT	TC	T GI	'A A'	T AC	F TC	T G	FT '	TAT	GAA	GAC	913
5	Al	a Le	eu Le	eu Ly	rs GI	lu Al	a Il	е ні	s Va	1 11	e Se	er Cy	rs G	ly '	Tyr	Glu	Asp	
		29						95					100					
	AA -	A AC	C G	VA TO	E CC	A AC	C GA	G CI	T GG	C TG	G AT	T TA	CG	<b>3C</b> 2	rœ	ATT	ACA	960
	ΓĀ	s Th	ur Gl	lu Tr	p G1			u Le	u Gl	y Tr	p Il	.е Ту	T G	ly s	Ser	Ile	Thr	
10	30		m > r	~ ~~			 					15					32	0
	GA	M GA	. T.	CTT	A AC	A GG	T TT	CAA	G AT	G CA	T TG	c cc	TG	₽¥.	T			1000
	GI	uns	b 11	e Le			y Ph	e ry	s Me			'S Ar	gr GI	-y				
					32	43				3	30							
15	(2	) IN	FORM	ATIO	N FO	R SE	Q ID	NO:	10:									
							ACTEI											
							333			ids								
							ino a											
20				(D)	TOPO	LOGY	: li	near										
		(1.	i) M	OLEC	JLE '	TYPE:	: per	otide	3									
							inte											
							UPTI											
25			₹ Va.	l Arg	Pro	Tha	Phe	Val	Lys	Glu	ı Arg	y Arg	, Al	a Ma	et	Lys	Arg	
	27					5	_				ıo					15		
	GIU	177	GI			e Lys	Val	Arg	_		1 Ala	Leu	ı Va	L A	la :	Lys	Ala	
30	G1 n	T		20				_	2						30			
	GIII	гус	3!		PIC	GIU	Gly			Met	Gln	Asp			מב ו	Pro	Trp	
	Pro	Glu			ጥኮተ	Tage	A ~~	4: u: c		<b>C1</b>	. 14-1			<b>4</b> 5			_	
		50	) )	1131	11.11	- Lys	Asp 55	_	PIO	GTĀ	Met	_		ı Va	<b>JT</b> 1	Phe	Leu	
5	Gly			Glv	Glv	His	Asp		Glu	G1v	, Acm		0		1		•	•
	65			1	2	70		•••	Olu	GLY	7		Let	ı PI	.O A	uy .	Leu 80	
	Val	Tyr	Val	Ser	Arg	Glu	Lys	Ara	Pro	Glv			Hic	: H1	e ī	are 1		•
					85	5	-	,		9			,	•••		95 95		
0	Ala	Gly	Ala	Met	Asn	Ala	Leu	Val	Arg			Gly	Val	Ιe	u T			
				100	t				105	_		_	-	_	.10			
	Ala	Pro	Phe	Met	Leu	Asn	Leu	Asp	Cys	Asp	His	Tyr	Leu			ısn S	Ser	
			115					120	)				12	5				
5	Lys	Ala	Val	Arg	Glu	Ala	Met	Cys	Phe	Leu	Met	Asp	Pro	Gl	n I	le G	31y	
		130					135					140	)					
	Arg	Lys	Val	Cys	Tyr		Gln	Phe	Pro	Gln	Arg	Phe	Asp	Gl	y I	le A	sp	
	145					150					155	5					160	
)	Arg	His	Asp	Arg	Tyr	Ala	Asn	Arg	Asn	Thr	Val	Phe	Phe	Ası	o I	le A	sn	
					165					170	)					175		

Met Lys Gly Leu Asp Gly Ile Gln Gly Pro Val Tyr Val Gly Thr Gly

	•	180	18	5	190
5		Phe Arg Arg G 195	in Ala Leu Tyr 200	Gly Tyr Glu	Pro Pro Lys Gly 205
	Pro Lys A 210	arg Pro Lys Me	et Val Thr Cys 215	Gly Cys Cys 22	Pro Cys Phe Gly
10	Arg Arg A 225		ys Lys His Ser 30	Lys Asp Gly 235	Gly Asm Ala Asm 240
	- 26				, - XX X X
		245		250	233
15	His Met A	sn Phe Glu Ly	s Lys Phe Gly	Gln Ser Ala	Ile Phe Val Thr
		260	26!		270
	Ser Thr L				Ser Ser Pro Ala
		275	280		285
20	Ala Leu L	eu Lys Glu Al	a Ile His Val	Ile Ser Cys	Gly Tyr Glu Asp
	290		295	30	0
	Lys Thr G	lu Trp Gly Se	er Glu Leu Gly	Trp Ile Tyr	Gly Ser Ile Thr
	305	3	10	315	320
25	Glu Asp I	le Leu Thr Gl	y Phe Lys Met	His Cys Arg	Gly
		325		330	
					**
30		MATION FOR SE			
	(i)	SEQUENCE CHAP			
			622 amino aci	ds	
		(B) TYPE: ar			
35		(D) TOPOLOGY			
		MOLECULE TYPE			
		FEATURE:	C-terminal fi	ragment	
	(IA)	(A) NAME/KE	7.		
40		(B) LOCATION			
			IFORMATION: Xa	a indicates (	alu or Ive
	(xi) :		RIPTION: SEQ 1		ord or Lys
45					Ala Met Lys Arg
	1	1_g 1.50 a.s. 5		10	15
			s Val Arm Ile		Val Ala Lys Ala
		20	25		30
50	Gln Lys Va				Gly Thr Pro Trp
		35	40	•	45
	Pro Gly As	sn Asn Thr Ly	s Asp His Pro	Gly Met Ile	Gln Val Phe Leu
	50	_	55	- 60	
<i>55</i>					

	G]	Ly G.	ln Se	er Gl	y Gl	y Hi	s Ası	o Thi	c Glu	ı Gly	/ Asn	Glu	Leu	Pro	An	g Leu
	Ć	5				7	O				7	<b>'</b> 5				80
5	Va	T L	YI Va	al Se	r An	g Glu	ı Lys	Arg	Pro	Gly	Phe	Leu	His	His	Ly:	s Lys
					8	35				9	0					95
	Al	a GI	ly Al	la Me	t Ası	n Ala	Leu	ı Val	. Arg	, Val	Ser	Gly	Val	Leu	Th	Asn
				10	Ю .				10	5				11	lO	
. 10	Al	a Pr	no Ph	ne Me	t Le	ı Ast	Leu	Asp	Cys	Asp	His	Tyr	Leu	Asn	Asr	Ser
			11	15				12	0				12	5		
	Ly	s Al	a Va	ıl Ar	g Glu	ı Ala	Met	Cys	Phe	Leu	Met	Asp	Pro	Gln	Ile	Gly
		13	<b>50</b>				135	5				140	C			
15	Ary	g Ly	s Va	l Cys	з Тут	· Val	Gln	Phe	Pro	Gln	Arg	Phe	Asp	Gly	Ile	Asp
	14:	)				150	)				155	5				160
	Arq	j Hi	s As	p Arg	J Tyr	Ala	Asn	Arg	Asn	Thr	Val	Phe	Phe	Asp	Ile	Asn
20					165	5				170	)				17	<b>'</b> 5
20	Met	Ly	s Gly	y Leu	ı Asp	Gly	Ile	Gln	Gly	Pro	Val	Tyr	Val	Gly	Thr	Gly
				180	)				185	5				19	0	
	Cys	va.	l Phe	e Arg	Arg	Gln	Ala	Leu	Tyr	Gly	Tyr	Glu	Pro	Pro	Lys	Gly
25			19	5				200	i				205	5		
	Pro	Lys	s Arg	J Pro	Lys	Met	Val	Thr	CAa	Gly	Cys	Cys	Pro	Cys	Phe	Gly
		210	J				215					220				
	Arg	Arg	Arg	, Lys	Asp	Lys	Lys	His	Ser	Lys	Asp (	Gly	Gly	Asn	Ala	Asn
30	225		<b></b>	_		230					235					240
	GIĀ	Leu	ı Ser	Leu	Glu	Ala	Ala	Xaa	Asp	Asp	Lys (	Glu :	Leu	Leu	Met	Ser
	772 -		_		245					250					25	5
	nis	Met	: Asn	Phe	GIu	Lys	Lys	Phe		Gln :	Ser A	Ala :	Ile 1	Phe '	Val	Thr
35	C	ma		260					265					270	)	
	ser	Thr	Leu	Met	Glu	Gln	Gly	Gly	Val	Pro 1	Pro S	Ser S	Ser S	Ser :	Pro	Ala
	λla	Lou	275		<b>~</b> 1			280			. 1		285			
	Ara	290	Leu	Lys	GIU	Ala .	Ile	His '	Val	Ile S	Ser C	ys (	ly 7	[ĀĽ (	Glu .	Asp
40	T				<b>~</b> 3	_	295					300				
	305	ш	GIU	Trp	GIĀ	Ser (	Glu 1	eu (	Sly :	Ltb 1	le T	yr c	Sly S	er 1	[leˈ	Thr
		۸	T1_	•	<b></b>	310					315					320
45	GLU	ASP	TTE	Leu	Thr (	GIY I	Phe I	ya v	let 1		ys A	rg G	ly 1	rp /	lrg !	Ser
	Tlo	m	<b>~</b>	<b>5</b> 4- 1	325	_				330					335	
	me	TYE	Cys	Met	Pro !	Lys I	eu F	to y	la F	he L	ys G	ly S	er A	la F	oré	[le
	λœn	T 011	C	340			_		3 <b>45</b>					350		
50	ASII	Leu	ser	Asp .	Arg I	Leu A	isn G	ln V	al I	eu A	rg T	rp A	la L	eu G	ly s	er
			355				;	360					365			
	Val (	etn G	116	Phe 1	Phe S	Ser H	is H	is C	ys P	ro A	la Ti	T P	yr G	ly P	he I	ys
	•	3/0				3	375				3	380				
55	Gly (	ътĀ	гÀЗ	Leu 1	lys 1	rp L	eu G	lu A	rg P	he Al	la Ty	r Va	al As	sn Ti	hr I	hr

	385					390	)				395	5				400
	Ile	Tyr	Pro	Phe	Thr	Ser	Leu	Pro	Leu	Leu	Ala	Tyr	Cys	Thr	Leu	Pro
		•			405					410		-	-		41	
5	*10	T10	~~	T ann			λ <b>~</b>	T	Dho			D	D	Ile		
	VTG	116	Cys			11111	vəb	гЛЭ			rec	PLO	PIO			1111
				420					425					43		
	Phe	Ala	Ser	Leu	Phe	Phe	Ile	Ala	Leu	Phe	Leu	Ser	Ile	Phe	Ala	Thr
10			435					440					44	5		
	Gly	Ile	Leu	Glu	Leu	Arg	Trp	Ser	Gly	Val	Ser	Ile	Glu	Glu	Trp	Trp
	Arm	Asn	Glu	Gln	Phe	Tro	Val	Ile	Glv	Glv	Ile	Ser	Ala	His	Leu	Phe
15	465					470			1	1	475					480
		1701	T3-0	C1-	C1			T	Val	T 011			T30	700	TTh-∞	
	ATS	var	тте	GIII	_		Leu	гÃ2	var			GTĀ	116	Asp		
					485		_			490					49	
20	Phe	Thr	Val	Thr	Ser	Lys	Ala	Thr	Asp	Asp	Glu	Glu	Phe	Gly	Glu	Leu
				500					505	•				51	0	
	Tyr	Thr	Phe	Lys	Trp	Thr	Thr	Leu	Leu	Ile	Pro	$\mathbf{Pro}$	Thr	Thr	Val	Leu
			515					520	)				52	5		
25	Ile	Ile	Asn	Leu	Val	Gly	Val	Val	Ala	Gly	Ile	Ser	Asp	Ala	Ile	Asn
20		530				_	535			_		540				
	Asn		Тулг	Gln	Ser	TTT			Teu	Phe	Glv			Phe	Phe	Ser
	545	0_1	-1-	<b>U</b>	-	550	_				555	_				560
30		<b>Т</b>	t/al	T10	v-1			Т+	D	Pho			Clar	Lou	Mat	
30	FIRE	тъ	vai	TIE		шѕ	Leu	ıĂr	PIO			пХ2	GLY	Leu		-
	_		_	_	565	_	_			570		_	_		57	
	Arg	GIN	Asn	_	Thr	Pro	Thr	TT6			He	Trp	Ser	Val		Leu
35				580					585	j				590	)	
33	Ala	Ser	Ile	Phe	Ser	Leu	Leu	Trp	Val	Arg	Ile	Asp	Pro	Phe	Val	Met
			595					600			1		605	5		
	Lys	Thr	Lys	Gly	Pro	Asp	Thr	Thr	Met	Cys	Gly	Ile	Asn	Cys		
10		610					615					620	)			
40				-												
	(2)	INFO	RMAT	'ION	FOR	SEQ	ID N	ю: 1	2:							
	•			UENC												
. =		\-,	_	A) LE												
45			-	-												
			•	3) T												
			•	) TC												
		-		ECUL												
50		(V)	FRAG	MENT	TYF	E: i	nter	mal	frag	ment	;					
		(xi)	SEC	UENC	E DE	SCRI	PTIC	N: S	EQ I	D NO	: 12	:				
	Gln	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Arg	Trp							
	1				5			-	-							
<i>55</i>	_				_											

	(2) INFORMATION FOR SEQ ID NO: 13:	
	(1) SEQUENCE CHARACTERISTICS:	
5	(A) LENGTH: 50 base pairs	
-	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
10	(ii) MOLECULE TYPE: other nucleic acid	
10	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 13:	
	GAGAGAGAG GAGAGAGAGA ACTAGTCTCG AGTTTTTTTT TTTTTTTTTT	50
15	(2) INFORMATION FOR SEQ ID NO: 14:	
	(1) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 13 base pairs	
	(B) TYPE: nucleic acid	
20	(C) STRANDEDNESS: double	•
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: other nucleic acid	
	(A) DESCRIPTION: /desc = "Synthetic DNA"	
25	(ix) FEATURE:	
	(A) NAME/KEY:	
	(B) LOCATION:14	
	(D) OTHER INFORMATION: single strand	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:	
	AATTOGGCAC GAG	13
	(2) INFORMATION FOR SEQ ID NO: 15:	
	(1) SEQUENCE CHARACTERISTICS:	
5	(A) LENGTH: 20 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
0	(ii) MOLECULE TYPE: other nucleic acid	
	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 15:	
	GACTGAAGAT AAGCCAAAAG	
5		20
	(2) INFORMATION FOR SEQ ID NO: 16:	
	(1) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 19 base pairs	
)	(B) TYPE: nucleic acid	

	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
5	(ii) MOLECULE TYPE: other nucleic acid	
	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 16:	
	GGAATGATGA ATTTGCCGG	19
10		
	(2) INFORMATION FOR SEQ ID NO: 17:	
	(A) herein. 20 base pairs	
15	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: other nucleic acid	
20	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:	
	TGCAGGCAAC TTTGGCATGC	20
	(2) INFORMATION FOR SEQ ID NO: 18:	
25	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 30 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
30	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: other nucleic acid	
	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:	
35	AGCAACACGA GCAAGATGAG GAGGATGACT	30
		00
	(2) INFORMATION FOR SEQ ID NO: 19:	
	(1) SEQUENCE CHARACTERISTICS:	
40	(A) LENGTH: 28 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
45	(ii) MOLECULE TYPE: other nucleic acid	
	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 19:	
	COGGATOCIT CAACCCITCT TOGATITC	28
50		
	(2) INFORMATION FOR SEQ ID NO: 20:	

	(1) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 29 base pairs	
5	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: other nucleic acid	
10	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(XL) SEQUENCE DESCRIPTION: SEQ ID NO: 20:	
	COGGATCCAC GGCAATGCAT CTTGAAACC	29
	(2) Титого стана	25
15	(2) INFORMATION FOR SEQ ID NO: 21:	
	(1) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 27 base pairs	
	(B) TYPE: nucleic acid	
20	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(11) MOLECULE TYPE: other nucleic acid	
	(A) DESCRIPTION: /desc = "Synthetic DNA"	
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21: GGTTAGCATA TTGTTTGTAG CATTGGG	
	SOTTABOATA TIGITIGIAG CATIGOG	27
	(2) INFORMATION FOR SEQ ID NO: 22:	
	(i) SEQUENCE CHARACTERISTICS:	
30	(A) LENGTH: 27 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
<i>35</i>	(ii) MOLECULE TYPE: other nucleic acid	
	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:	
	ATCAATGAAA TATGTATAGT TCATAGC	
40		27
	(2) INFORMATION FOR SEQ ID NO: 23:	
	(1) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 27 base pairs	
45	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: other nucleic acid	
50	(A) DESCRIPTION: /desc = "Synthetic DNA"	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 23:	

#### CITICGITCT TITGGITTIG CCATGGC

27

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(11) MOLECULE TYPE: other pucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

AGACTITITA CAAACAAGAT AAATCCC

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#### Claims

1. A DNA coding for any one of the following proteins (A) to (C):

(A) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 2 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 2;

(B) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 4 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 4; and

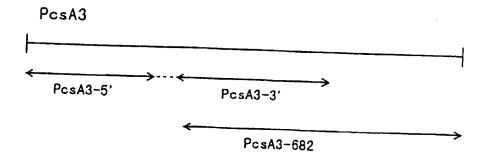
(C) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 8 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 8, and an amino acid sequence shown in SEQ ID NO: 11 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 11.

- 2. A recombinant vector comprising all or a part of the DNA as defined in claim 1.
- A transformed cell transformed with the DNA as defined in claim 1.

4. A method for controlling cellulose synthesis in a cell, comprising the steps of introducing the DNA as defined in claim 1 into the cell, and expressing RNA having a nucleotide sequence homologous to the DNA as defined in claim 1 or a nucleotide sequence complementary to the DNA as defined in claim 1.

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# FIG. 1

SEQ ID NO: 14

5' AATTCGGCACGAG 3'
3' GCCGTGCTC 5'---

FIG. 2

	10	20	30	40	50	60
PcsA3-682	CCGACATTCGTGA	AGGAGCGTCG	AGCTATGAAG	AGAGAATATG	AAGAATTCAA	GGTTAGG
(SEQ ID NO: 5)	:::::::::::::::::::::::::::::::::::::::	:::::::::	:::::::::	:::::::::	::::::::::	::::::
PcsA3-3'	CCGACATTCGTGA	AGGAGCGTCG	AGCTATGAAG	AGAGAATATG	AAGAATTCAA	GGTTAGG
(SEQ ID NO: 9)	20	30	40	50	60	70
	70	80	90	100	110	120
PcsA3-682	ATAAATGCACTTG	TAGCCAAAGC	CAAAAGGTT	CCTCCAGAAG	GGTGGATCAT	GCAAGAT
			:::::::::	::::::::::	111111111	
	-LEALARDOLOTTO	TACOCIAACO	TTODALALACE	COLCOAGAAA	CCTCCATCAT	GGAAGAT
, , , , , , , , , , , , , , , , , , ,	130	140	150	160	170	180
PcsA3-682	GGGACACCATGGC	CAGGAAACAA	TACTAAAGAT	CACCCTGGTA	TGATTCAAGT	ATTTCTC
	:::::::::::::::	::::::::::	:::::::::	::::::::::		1111111
PcsA3-3'	GGGACACCATGGC	CAGGAAACAA	TACTAAAGAT	CACCCTGGTA	TGATTCAAGT	ATTTCTC
,	140	150	160	170	180	190
	190	200	210	220	230	240
PcsA3-682	GGTCAAAGTGGAG					
1 00/10 002		::::::::::				
PcsA3-3'	GGTCAAAGTGGAG				CGTCTCGTCTA	TGTATCT
. 55/15	200	210	220	230	240	250
	250	260	270	280	290	300
PcsA3-682	CGAGAGAAAAGGC					
100/10 002		*******				
PcsA3-3'	CGAGAGAAAAGGC	CAGGTTTCTT	GCATCACAAG	AAAGCTGGTG	CCATGAACGC	CCTTGTT
, 50,15	260	270	280	290	300	310
	310	320	330	340	350	360
PcsA3-682	CGGGTCTCGGGGG			• • • •		
. 50,15 552	::*::::::::					:::*:::
PcsA3-3'	CGTGTCTCGGGGG	TGCTTACAAA	TGCTCCTTTT	ATGTTGAACT	TGGATTGTGA	CCACTAT
	320	330	340	350	360	370
	370	380	390	400	410	420
PcsA3-682	TTAAATAACAGCA	AGGCTGTAAG	AGAGGCTATO	TGTTTCTTG	ATGGACCCTCA	AATTGGA
	:::::::::::::::::::::::::::::::::::::::	:::::::::::	:::::::::		:::::::::::::	::::::::
PcsA3-3	TTAAATAACAGCA	AGGCTGTAAG	AGAGGCTATO	TGTTTCTTG	ATGGACCCTC/	AATTGGA
	380	390	400	410	420	430
	430	440	450	460	470	480
PcsA3-682	AGAAAGGTTTGCT	ATGTCCAATT	CCCTCAACG	TTCGATGGT	ATTGATAGACA	ATGATCGA
	:::::::::::::::::::::::::::::::::::::::	:::::::::::	:::::::::			
PcsA3-3'	AGAAAGGTTTGC1					
	440	450	460	470	480	490
	490	500	510	520	530	540
PcsA3-682	TATGCCAATCGG				•••	•
. 53/10 002	:::::::::::::::			_		
PesA3-3'	TATGCCAATCGG			_		
r Cano 1	500	510	520	530	540	550
	500	3.0	520	300	3-0	330

FIG. 3

	550	560	570	580	590	600
PcsA3-682	GGCCCTGTATAT	GTCGGCACGG	GGTGTGTTTT	CAGAAGGCAA	GCTCTTTATG	GTTATGAA
SEQ ID NO: 5)	::::::::::	:::::::::::	::::::::	::::::::::	:::::::::	:::::::
PcsA3-3'	GGCCCTGTATAT	GTCGGCACGG	GGTGTGTTTT	CAGAAGGCAA	GCTCTTTATG	GTTATGAA
SEQ (D NO: 9)	560	570	580	590	600	610
	610	620	630	640	650	660
PcsA3-682	CCTCCAAAGGGA	CCTAAGCGCC	CGAAAATGGT.	AACCTGTGGT	TGCTGCCCTT	GTTTTGGA
	::::::::::	::::::::	::::::::	:::::::::	::::::::::	:*:::::
PcsA3-3'	CCTCCAAAGGGA	CCTAAGCGCC	CGAAAATGGT	AACCTGTGGT	TGCTGCCCTT	GCTTTGGA
	620	630	640	650	660	670
	670	680	690	700	710	720
PcsA3-682	CGCCGCAGAAAG	GACAAAAAGC	ACTCTAAGGA	TGGTGGAAAT	GCAAATGGTC	TAAGCCTA
	:::::::::::	:::::::::	:::::::::::::::::::::::::::::::::::::::	:::::::::	::::::::	:::::::
PcsA3-3'	CGCCGCAGAAAG	GACAAAAAGC	ACTOTAAGGA	TGGTGGAAAT	GCAAATGGTC	TAAGCCTA
	680	690	700	710	720	730
	730	740	750	760	770	780
PcsA3-682	GAAGCAGCCAAA	GATGACAAGG	AGTTATTGAT	GTCCCACATG	AACTTTGAAA	AGAAATTT
	:::::::::*::	::::::::	:::::::::	:::::::::::::::::::::::::::::::::::::::	::::::::	:::::::
PcsA3-3'	GAAGCAGCCGAA	GATGACAAGG	AGTTATTGAT	GTCCCACATG	AACTTTGAAA	AGAAATTT
	740	750	760	770	780	790
	790	800	810	820	830	840
PcsA3-682	GGACAATCAGCC	ATTTTTGTAA	CTTCAACACT	GATGGAACAA	GGTGGTGTCC	CTCCTTCT
	:::::::::::::::::::::::::::::::::::::::	::::::::	::::::::	:::::::::	::::::::	:::::::
PcsA3-3'	GGACAATCAGCC	ATTTTTGTAA	CTTCAACACT	GATGGAACAA	GGTGGTGTCC	CTCCTTCT
	800	810	820	830	840	850
	850	860	870	880	890	900
PcsA3-682	TCAAGCCCCGCA	GCTTTGCTCA	AAGAAGCCAT	TCATGTAATT	AGTTGTGGTT.	ATGAAGAC
	::::::::::					
PcsA3-3'	TCAAGCCCTGCA	GCTTTGCTCA	AAGAAGCCAT	TCATGTAATT	AGTTGTGGTT.	ATGAAGAC
	860	870	880	890	900	910
	910	920	930	940	950	960
PcsA3-682	AAAACAGAATGG	GGAAGCGAGC	TTGGCTGGAT	TTACGGCTCG	ATTACAGAAG	ATATCTTA
	:::::*:::::					
PcsA3-3'	AAAACCGAATGG	GGAAGCGAGC	TTGGCTGGAT	TTACGGCTCG	ATTACAGAAG	ATATCTTA
	920	930	940	950	960	970
	970	980				
PcsA3-682	ACAGGATTCAAG	ATGCATTGCC	GTGGAT			
	:::::*:::::	::::::::	:::::			
PcsA3-3	ACAGGTTTCAAG	ATGCATTGCC	GTGGAT			
	990	aan	1000			

FIG. 4

		90 1845 1800
		€.± . *

# Europäisches Patentamt Europ an Patent Office Office européen des brevets



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## (54) Cellulose synthase gene

(57) .mRNA was extracted at the stage for cotton plant fibrous cells to accumulate cellulose, and cDNA's complementary thereto were synthesized to construct a cDNA library. Clones of a number of 750 were arbitrarily selected from the library, and they were randomly subjected from to sequencing. Those having homology to

an amino acid sequence deduced from a gene of cellulose 4-β-glucosyltransferase (bcsA) of cellulose synthase operon of acetic acid bacterium were selected from obtained nucleotide sequences of the respective clones. Thus, DNA coding for cellulose synthase was obtained.



# **EUROPEAN SEARCH REPORT**

Application Number EP 98 30 2489

	DOCUMENTS CONSIL	DERED TO BE RELEVANT			
Category	Citation of document with of relevant pas	indication, where appropriate, sages	Relevant to plaim	CLASSIFICATION OF THE APPLICATION (Int.CI.6)	]
рх	WO 98 00549 A (THE	AUSTRALIAN NATIONAL	1-4	C12N15/54	
X,D	* page 1, line 3 - * page 2, line 21 * example 8 * 'Sequence Listing: PEAR, J.R. ETAL.: homologs of the ba encoding the catal cellulose synthase PROC.NATL.ACAD.SCI vol. 93, October 19	line 11 * - page 7, line 28 * - SEQ ID NO.9 and 10 "Higher plants contain cterial celA genes ytic subunit of	1-4		
Y	xP002061424 * the whole document WO 91 13988 A (THE UNIVERSITY OF TEXAS 19 September 1991 * page 1, line 18 * page 5, line 15 * figure 1; example	BOARD OF REGENTS, THE SYSTEM) - line 26 * - page 8, line 13 *	1-4	TECHNICAL FIELDS SEARCHED (Int.Cl.6)	
	cotton fiber" PLANT PHYSIOLOGY,	Glucan synthesis in the 93, pages 1149-1156,	1-4	·	
	WO 98 18949 A (CALG * page 7, line 14 - * figures 3,6,8; ex		1-4		
	The present search report has	peen drawn up for ail slaims			
	Prace of searon	Date of completion of the search		Examiner	
ſ	MUNICH	8 December 1998	Dona	ath, C	
X partici Y partici docum A technic O : non-w	TEGORY OF CITED DOCUMENTS usually relevant if taken alone usually relevant if combined with anothern of the same category ological background written disclosure sedate document	T theory or principle E earlier patent document after D document after in L document after in 3 member of the san cocument	ment, but publish the application other reasons		



Application Number

EP 98 30 2489

CLAIMS INCURRING FEES
The present European patent application comprised at the time of filing more than ten claims.
Only part of the claims have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims and for those claims for which claims fees have been paid, namely claim(s):
No claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims.
LACK OF UNITY OF INVENTION
The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:
see sheet 8
All further search fees have been paid within the fixed time limit. The present European search report has been drawn up for all claims.
As all searchable claims could be searched without effort justifying an additional fee, the Search Division did not invite payment of any additional fee.
Only part of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the inventions in respect of which search fees have been paid, namely claims:
None of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the invention first mentioned in the claims, namely claims:
1-4 (partially)
; ;



# LACK OF UNITY OF INVENTION SHEET B

Application Number

EP 98 30 2489

The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

1. Claims: 1-4 (partially)

acid sequence shown in SEQ ID NO:2 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO:2, a recombinant vector comprising all or part of said DNA, a cell being transformed with said DNA, and a method for controlling cellulose synthesis in a cell by the use of said DNA.

2. Claims: 1-4 (partially)

Claims 1-4 (partially) refer to a DNA coding for a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO:4 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO:4, a recombinant vector comprising all or part of said DNA, a cell being transformed with said DNA, and a method for controlling cellulose synthesis in a cell by the use of said DNA.

3. Claims: 1-4 (partially)

Claims 1-4 (partially) refer to a DNA coding for a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO:8 and in SEQ ID NO:11 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO:8 and/or SEQ ID NO:11, a recombinant vector comprising all or part of said DNA, a cell being transformed with said DNA, and a method for controlling cellulose synthesis in a cell by the use of said DNA.

#### SEQ ID NO: 6 for PcsA3.

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PcsA1 is different from CelA1 reported by Pear et al. (Proceeding of National Academy of Science, USA (1996), 93, 12637-12642) in nucleotide sequence by 28 nucleotides. As a result, the former is different from the latter in amino acid sequence encoded thereby by 10 amino acid residues. In general, the sugar chain specificity and the substrate specificity of the sugar chain transferase are extremely changed by point mutation of the nucleotide of DNA (Yamamoto and Hakomori, The Journal of Biological Chemistry (1990) 265, 19257-19262). Therefore, it is unclear whether or not CelA1 codes for a protein having the cellulose synthase activity. Incidentally, the 48th Arg, the 56th Ser, the 81st Asn, the 104th Ala, the 110th Ser, the 247th Asp, the 376th Asp, the 386th Ser, the 409th Arg, and the 649th Ser in the amino acid sequence encoded by CelAl correspond to Gln, Ile, Ser, Thr, Pro, Asn, Glu, Pro, His, and Gly in PcsA1 respectively.

PcsA2 of the present invention contains the same sequence as that of CelA2 reported by Pear et al. However, CelA2 has an incomplete length, and it does not contain the entire coding region. CelA2 corresponds to nucleotide numbers of 1083 to 3311 in the nucleotide sequence of PcsA2 shown in SEQ ID NO: 3.

Any of the amino acid sequences shown in SEQ ID NOs: 2, 4, 6, 8, 10, and 11 is a novel sequence. All genes having nucleotide sequences coding for the amino acid sequences are included in the present invention.

The amino acid sequences described above may include deletion, substitution, insertion, and/or addition of one or more amino acid residues provided that the characteristic of the gene of the present invention is not substantially affected. The deletion, substitution, insertion, and/or addition of one or more amino acid residues as described above is obtainable by modifying the DNA's coding for the amino acid sequences shown in SEQ ID NOs: 2, 4, 6, 8, 10, and 11 randomly in accordance with the ordinary mutation treatment or intentionally in accordance with the site-directed mutagenesis method. As described above, in general, the sugar chain specificity and the substrate specificity of the sugar chain transferase are extremely changed by point mutation of the nucleotide of DNA. Therefore, DNA coding for a protein having the cellulose synthase activity is selected from the modified DNA's. The cellulose synthase activity can be measured, for example, by means of the method described by T. Hayashi: Measuring-β-glucan deposition in plant cell walls. in Modern Methods of Plant Analysis: Plant Fibers, eds. H. F. Linskens and J. F. Jackson, Springer-Verlag, 10: 138-160 (1989).

Those harboring proteins or genes partially different from the sequences shown in Sequence Listing may exist depending on, for example, the variety of cotton plant or natural mutation. However, such genes are also included in the gene of the present invention. Such a gene may be obtained as DNA which is hybridizable under the stringent condition with all or a part of the coding region of the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, or 9. The "stringent condition" referred to herein indicates a condition under which a so-called specific hybrid is formed, and non-specific hybrid is not formed. It is difficult to definitely express such a condition by using a numerical value. However, for example, the stringent condition is exemplified by a condition under which nucleic acids having high homology, for example, DNA's having homology of not less than 80 % undergo hybridization with each other, and nucleic acids having homology lower than the above do not undergo hybridization with each other.

# <5> Utilization of gene of the present invention

The DNA of the present invention makes it possible to control the cellulose synthesis in prokaryotic cells such as acetobacterium and/or eukaryotic cells such as yeasts belonging to, for example, the genus <u>Saccharomyces</u>, cells of plant such as cotton plant, and cultured cells of mammals and the like.

Specifically, the cellulose synthesis in the cells as described above can be facilitated, for example, by connecting a promoter to an upstream region of the DNA of the present invention, inserting an obtained fragment into an appropriate vector to construct a recombinant vector, and introducing the vector into the cells. Alternatively, the cellulose synthesis in the cells can be suppressed by introducing an antisense gene of the DNA of the present invention into the cells.

The promoter and the vector may be selected from those ordinarily utilized to express heterogeneous genes, and the method ordinarily employed to express heterogeneous genes may be used as the transformation method. Specifically, in the case of yeast, it is possible to use a protein-expressing kit produced by Invitrogen, i.e., Pichia Expression Kit, and a vector pPIC9 contained in this kit. For example, COS7 cells may be used as mammalian cultured cells, and a vector CDM8 may be used therefor.

The present invention provides the DNA coding for cellulose synthase. The DNA provides a new method for controlling cellulose production by incorporating the DNA into prokaryotic cells and eukaryotic cells.

# Brief Description of the Drawings

Fig. 1 shows a relationship between two clones of PcsA3 as an embodiment of the DNA of the present invention. Regions interposed between arrows indicate regions for which nucleotide sequences have been determined. A dotted line indicates a region for which no nucleotide sequence has been determined.

- Fig. 2 shows a structure of EcoRI adapter.
- Fig. 3 shows comparison between sequences of PcsA3-682 and PcsA3-3' (former half).
- Fig. 4 shows comparison between sequences of PcsA3-682 and PcsA3-3' (latter half). ":" indicates coincident nucleotides, and "\*" indicates non-coincident nucleotides.

# Best Mode for Carrying Out the Invention

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Examples of the present invention will be explained below.

#### <1> Preparation of total RNA from cotton plant

Cotton plant (Gossypium hirsutum L.) Coker 312 was used as a material. Fiber cells on 16 to 18 days post anthesis

which 375 mg of DTT as a powder was added, followed by addition of 200 ml of XT buffer (obtained by adjusting 0.2 M sodium borate containing 30 mM EDTA and 1 % SDS to be pH 9.0, and then applying a diethylpyrocarbonate treatment, followed by autoclaving to obtain a solution to which vanadylribonucleoside was added to give a concentration of 10 mM) having been heated to 90 to 95 °C. An obtained solution was sufficiently agitated.

The solution was added with 100 mg of protease K, and it was agitated again. The solution was incubated at 40 °C for 2 hours, and then it was added with 16 ml of 2 M KCl. The solution was sufficiently agitated again, and it was left to stationarily stand in ice for 1 hour, followed by centrifugation for 20 minutes (4 °C) at 12,000 g by using a high speed refrigerated centrifuge.

An obtained supernatant was filtrated, and floating matters were removed. The solution was transferred to a measuring cylinder to measure the volume. The solution was transferred to another centrifuge tube, to which lithium chloride was added in an amount of 85 mg per 1 ml of the extract solution to give a final concentration of 2 M. The solution was left to stationarily stand at 4 °C overnight, and then precipitated RNA was separated by centrifugation for 20 minutes at 12,000 g. An obtained precipitate of RNA was washed and precipitated twice with cooled 2 M lithium chloride.

The obtained RNA was dissolved in 10 mM Tris buffer (pH 7.5) to give a concentration of about 2 mg/ml, to which 5 M potassium acetate was added to give a concentration of 200 mM. Ethanol was added thereto to give a concentration of 70 %, followed by cooling at -80 °C for 10 minutes. Centrifugation was performed at 4 °C for 10 minutes at 15,000 rpm, and then an obtained precipitate was suspended in an appropriate amount of sterilized water to give an RNA sample. As a result of quantitative measurement for the RNA sample, total RNA was obtained in an amount of 2 mg.

#### <2> Purification of mRNA

mRNA was purified as a poly(A)+ RNA fraction from the total RNA obtained as described above. Purification was performed by using Oligotex-dT30 <Super> (purchased from Toyobo) as oligo(dT)-immobilized latex for poly(A)+ RNA purification.

Elution buffer (10 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.1 % SDS) was added to a solution containing 1 mg of the total RNA to give a total volume of 1 ml, to which 1 ml of Oligotex-dT30 <Super> was added, followed by heating at 65 °C for 5 minutes and quick cooling on ice for 3 minutes. The obtained solution was added with 0.2 ml of 5 M NaCl, and it was incubated at 37 °C for 10 minutes, followed by centrifugation at 15,000 rpm for 3 minutes. After that, a supernatant was carefully removed.

An obtained pellet was suspended in 2.5 ml of Washing Buffer (10 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.5 M NaCl, 0.1 % SDS), and the suspension was centrifuged at 15,000 rpm for 3 minutes. After that, a supernatant was carefully removed. An obtained pellet was suspended in 1 ml of TE Buffer, and then it was heated at 65 °C 5 minutes. The suspension was quickly cooled on ice for 3 minutes, and then it was centrifuged at 15,000 rpm for 3 minutes to recover poly(A)+ mRNA contained in an obtained supernatant.

Thus, the poly(A)+ mRNA in an amount of about 10 μg was obtained from 1 mg of the total RNA. An aliquot of 5 μg thereof was used to prepare a cDNA library.

#### <3> Preparation of cDNA library

# (1) Synthesis of cDNA

The mRNA obtained as described above was used as a template to synthesis cDNA by using a  $\lambda$ ZAP cDNA synthesis kit produced by Stratagene. The following solution was prepared and mixed in a tube.

5.0  $\mu$ l 10 x 1st Strand Buffer (buffer for reverse transcription reaction); 3.0  $\mu$ l 10 mM 1st Strand Methyl Nucleotide Mix (5-methyl dCTP, dATP, dGTP, dTTP mixture); 2.0  $\mu$ l Linker-Primer (linker and primer); H<sub>2</sub>O (adjusted to give a total volume of 50  $\mu$ l);

The respective components described above were contents of the kit. Linker-Primer had a sequence as shown in SEQ ID NO: 13. Methylated nucleotide was used because it was intended not to allow cDNA to be digested by the restriction enzyme reaction performed later on. The reaction solution was agitated well, and then 5.0 µg of poly(A)+ mRNA was added thereto, followed by being left to stand at room temperature for 10 minutes. Further, 2.5 µl of M-MuLV RTase (reverse transcriptase) was added (at this time, the total volume was 50 µl). The reaction solution was gently mixed, followed by centrifugation under a mild condition to allow the reaction solution to fall to the bottom of the tube. The reaction was performed at 37 °C for 60 minutes.

Next, the following solution was prepared and mixed in the tube in a certain order.

45.0  $\mu$ l reaction solution containing cDNA primary chain; 40.0  $\mu$ l 10 x 2nd Strand Buffer (buffer for polymerase reaction); 6.0  $\mu$ l 2nd Strand Nucleotide Mixture (A, G, C, T mixture); 302.0  $\mu$ l H<sub>2</sub>O.

The following solution was further added. However, in order to allow RNase and DNA polymerase to simultaneously act, enzyme solutions were allowed to adhere to the wall of the tube. After that, a vortex treatment was promptly performed, and the reaction solutions were allowed to fall to the bottom of the tube by means of centrifugation to perform a reaction for synthesizing cDNA second strand at 16 °C for 150 minutes.

0.8 μl RNase H (RNA-degrading enzyme); 7.5 μl DNA polymerase I (10.0 u/μl).

1.0 μl RNase Block II (RNase inhibitor).

The reaction solution was added with 400 µl of a mixed solution of phenol: chloroform (1:1). Agitation was performed well, followed by centrifugation at room temperature for 2 minutes. An obtained supernatant was added with 400 µl of phenol: chloroform again, which was subjected to a vortex treatment and centrifugation at room temperature for 2 minutes. An obtained supernatant was added with the following solution to precipitate cDNA.

 $33.3 \,\mu$ l  $3 \,M$  sodium acetate solution;  $867.0 \,\mu$ l  $100 \,\%$  ethanol.

The obtained solution was left to stand at -20 °C overnight, and it was centrifuged at room temperature for 60 minutes. After that, washing was gently performed with 80 % ethanol, followed by centrifugation for 2 minutes. A supernatant was removed. An obtained pellet was dried, and it was dissolved in 43.5  $\mu$ l of sterilized water. An aliquot (39.0  $\mu$ l) was added with the following solution to blunt-end cDNA terminals.

5.0  $\mu$ l 10 x T4 DNA Polymerase Buffer (buffer for T4 polymerase reaction); 2.5  $\mu$ l 2.5 mM dNTP Mix (A, G, C, T mixture); 3.5  $\mu$ l T4 DNA polymerase (2.9  $u/\mu$ l).

The reaction was performed at 37 °C for 30 minutes, to which 50  $\mu$ l of distilled water was added, and then 100  $\mu$ l of phenol: chloroform was added thereto, followed by a vortex treatment and centrifugation for 2 minutes. An obtained supernatant was added with 100  $\mu$ l of chloroform, which was subjected to a vortex treatment, followed by centrifugation for 2 minutes. The supernatant was added with the following solution to precipitate cDNA.

 $7.0 \mu l$  3 M sodium acetate solution;  $226 \mu l$  100 % ethanol.

The solution was left to stand on ice for 30 minutes or more, and it was centrifuged at 4 °C for 60 minutes. An obtained precipitate was washed with 150 µl of 80 % ethanol, followed by centrifugation for 2 minutes and drying. The cDNA pellet was dissolved in 7.0 µl of EcoRl Adaptor solution, to which the following solution was added to ligate the EcoRl adapter to both ends of the cDNA. Sequences of respective strands of the EcoRl adapter are shown in SEQ ID NO: 14 and Fig. 2.

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- 1.0 μl 10 x Ligation Buffer (buffer for ligase reaction);
- 1.0 ul 10 mM ATP;

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- 1.0 µl T4 DNA ligase.
- The reaction solution was centrifuged under a mild condition, and it was left to stand at 4 °C overnight or more. The solution was treated at 70 °C for 30 minutes, and then it was centrifuged under a mild condition, followed by being left to stand at room temperature for 5 minutes. The reaction solution was added with the following solution to phosphorylate 5'-terminals of the EcoRl adapter.
- 10 1.0 μl 10 x Ligation Buffer (buffer for ligase reaction);
  - 2.0 µl 10 mM ATP;
- The reaction was performed at 37 °C for 30 minutes, followed by a treatment at 70 °C for 30 minutes. The solution was centrifuged under a mild condition, and it was left to stand at room temperature for 5 minutes. The following solution was further added thereto to perform a reaction at 37 °C for 90 minutes so that the Xhol site introduced by Linker-Primer was digested with Xhol, followed by being left to stand at room temperature to perform cooling.
- 20 28.0 μl <u>Xho</u>l Buffer;3.0 μl <u>Xho</u>l (45 u/μl).

The reaction solution was added with 5.0 µl of 10 x STE (10 mM Tris-HCl (pH 8.0), 100 mM NaCl, 1 mM EDTA), which was added into a centrifuge column for removing short fragments (Sephacryl Spin Column) to perform centrifugation at 600 g for 2 minutes to obtain an eluent which was designated as Fraction 1. This operation was further repeated three times to obtain Fractions 2, 3, and 4 respectively. Fractions 3 and 4 were combined, to which phenol: chloroform (1:1) was added and agitated well, followed by centrifugation at room temperature for 2 minutes. An obtained supernatant was added with an equal amount of chloroform, and an obtained mixture was agitated well. The mixture was centrifuged at room temperature for 2 minutes to obtain a supernatant to which a two-fold amount of 100 % ethanol was added, followed by being left to stand at - 20 °C overnight. The solution was centrifuged at 4 °C for 60 minutes, followed by washing with an equal amount of 80 % ethanol. The solution was centrifuged at 4 °C for 60 minutes to obtain a cDNA pellet which was suspended in 10 µl of sterilized water.

## (2) Preparation of cDNA library

The double strand cDNA obtained as described above was ligated with  $\lambda$  phage expression vector to prepare a recombinant vector. The following solution was prepared and mixed in a tube to perform a reaction at 12 °C overnight, followed by being left to stand at room temperature for 2 hours to ligate cDNA with the vector.

- 2.5 μl cDNA solution;
  - 0.5 µl 10 x Ligation Buffer;
  - 0.5 μl 10 mM ATP;
  - 1.0 μl λZAP vector DNA (1 μg/μl);
  - 0.5 μl T4 DNA ligase (4 Weiss u/μl).

#### (3) Packaging of phage DNA into phage particles

The phage vector containing the cDNA was packaged into phage particles by using an <u>in vitro</u> packaging kit (Gigapack II Gold packaging extract: produced by Stratagene). The recombinant phage solution was added to Freeze/ Thaw extract immediately after dissolution, and the solution was placed on ice, to which 15 µl of Sonic extract was added to perform mixing well by pipetting. The reaction solution was centrifuged under a mild condition, and it was left to stand at room temperature (22 °C) for 2 hours. The reaction solution was added with 500 µl of Phage Dilution Buffer, to which 20 µl of chloroform was further added, followed by mixing. In order to measure the titer of the library, an aliquot (2 µl) of 500 µl of the aqueous phase was diluted in a ratio of 1:10 with 18 µl of SM buffer (5.8 g of NaCl, 2 g of MgSo<sub>4</sub>\*7H<sub>2</sub>O, 50 ml of 1 M Tris-HCl (pH 7.5), and 5 ml of 2 % gelatin in 1 L). The diluted solution (1 µl) and the phage stock solution (1 µl) were plated respectively together with 200 p1 of a culture solution of <u>Escherichia coli</u> PLK-F' strain having been cultivated to arrive at a value of OD<sub>600</sub> of 0.5. That is, <u>Escherichia coli</u> PLK-F' strain was mixed with the phage solution to perform cultivation at 37 °C for 15 minutes. The obtained culture was added to 2 to 3 ml of top agar

(48 °C), which was immediately overlaid on NZY agar plate having been warmed at 37 °C. Cultivation was performed overnight at 37 °C, and appeared plaques were counted to calculate the titer. As a result, the titer was 1.2 x 106 pfu/ml.

#### (4) Amplification of library

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A centrifuge tube was added with the packaging solution containing about 50.000 recombinant bacteriophages and 600 µl of a culture solution of Escherichia coli PLK-F¹ strain having been cultivated to have a value of OD<sub>600</sub> of 0.5, followed by cultivation at 37 °C for 15 minutes. The culture solution was added with 6.5 ml of top agar having been maintained at 48 °C after dissolution, which was overlaid on 150 mm NZY plate having been warmed at about 37 °C, followed by cultivation at 37 °C for 5 to 8 hours. The respective plates were added with 10 ml of SM Buffer to perform cultivation at 4 °C overnight with gentle shaking. SM Buffer in the respective plates was collected in a sterilized polypropylene tube. The respective plates were rinsed with 2 ml of SM Buffer, and the rinsing solutions were collected in the same tube. Chloroform in an amount corresponding to 5 % of the total amount was added and mixed, followed by being left to stand at room temperature for 15 minutes. Bacterial cells were removed by centrifugation at 4,000 g for 5 minutes. An obtained supernatant was added with chloroform in an amount corresponding to 0.3 % of the total amount, and it was stored at 4 °C. The titer of the library amplified as described above was measured in the same manner as described above. As a result, the titer was 2.3 x 109 pfu/ml.

#### (5) Excision of plasmid from phage DNA

<u>In vivo</u> excision of the plasmid portion from the recombinant phage DNA was performed. The following solution was mixed in 50 ml of a conical tube to cause infection at 37 °C for 15 minutes:

culture solution of Escherichia coli XL1-Blue (OD<sub>600</sub> = 0.1) 200  $\mu$ l: phage solution after amplification 200  $\mu$ l (> 1 x 10<sup>5</sup> phage particles); helper phage R408 1  $\mu$ l (> 1 x 10<sup>6</sup> pfu/ml).

The mixed solution was added with 5 ml of 2 x YT medium to perform cultivation at 37 °C for 3 hours with shaking. A heat treatment was applied thereto at 70 °C for 20 minutes, followed by centrifugation at 4,000 g for 5 minutes. An obtained supernatant was decanted and transferred to a sterilized tube. Centrifugation was performed to obtain a supernatant which was diluted 100 times to obtain a solution. An aliquot (20  $\mu$ l) of the solution was mixed with 200  $\mu$ l of a culture solution of Escherichia coli XL1-Blue having been cultivated to obtain a value of OD<sub>600</sub> of 1.0 to cause infection at 37 ° C for 15 minutes. Aliquots (1 to 100  $\mu$ l) of the culture solution were plated on LB plates containing ampicillin, followed by cultivation at 37 °C overnight. Appeared colonies were randomly selected. Selected colonies were added with glycerol, and they were stored at -80 °C.

#### (6) Preparation of plasmid

Plasmids were prepared by using Magic Mini-prep kit produced by Promega. The culture fluid of <u>Escherichia coli</u> harboring the plasmid having been stored at -80 °C was inoculated into 5 ml of 2 x YT medium, followed by cultivation at 37 °C overnight. Centrifugation was performed for 5 minutes (4,000 rpm, 4 °C), and a supernatant was removed by decantation. An obtained bacterial cell pellet was added with 1 ml of TE buffer, followed by a vortex treatment. An obtained bacterial cell suspension was transferred to an Eppendorf tube, followed by centrifugation for 5 minutes (5,000 rpm, 4 °C). A resultant supernatant was removed by decantation.

An obtained bacterial cell pellet was added with 300  $\mu$ l of Cell Resuspension Solution, and it was sufficiently suspended therein. An obtained suspension was transferred to an Eppendorf tube. The suspension was agitated for 2 minutes with a mixer, to which 300  $\mu$ l of Cell Lysis Solution was added, followed by agitation until the suspension became transparent. Neutralization Solution (300  $\mu$ l) was added thereto, and agitation was performed by shaking with the hand, followed by centrifugation for 10 minutes (15,000 rpm).

Only an obtained supernatant was transferred to a new Eppendorf tube (1.5 ml). A suction tube was prepared, to which a cock, a miniature column and a syringe (injector) were connected in this order. A resin in an amount of 1 ml was charged into the syringe. The supernatant was poured into the syringe, and agitation was performed well, followed by suction. Column Washing Solution in an amount of 2 ml was added, and washing was performed while performing suction. Suction was continued for 1 to 2 minutes in order to dry up. The miniature column was removed from the equipment, and it was set in a new Eppendorf tube (1.5 ml). Sterilized water in an amount of 100 µl having been warmed at 65 to 70 °C was poured into the miniature column, and the column and the Eppendorf tube were centrifuged together for 1 minute (5,000 rpm). An eluted solution was transferred to an Eppendorf tube, to which 5 µl of 3 M sodium acetate aqueous solution was added, and 250 µl of cold ethanol was added thereto. The solution was centrifuged (15,000 rpm,

25 minutes), and a supernatant was discarded. An obtained precipitate was added with 1 ml of 70 % ethanol, followed by centrifugation again (15,000 rpm, 3 minutes). Ethanol was completely removed, and the tube was vacuum-dried in a desiccator. The precipitate was sufficiently dissolved in 20 μl of sterilized water, and an obtained solution was stored at -20 °C. An aliquot (1 μl) of the solution was dispensed, and it was subjected to electrophoresis together with volume markers to quantitatively determine the plasmid DNA.

### <4> Determination of nucleotide sequence of cDNA and homology search with gene data base

### (1) Determination of nucleotide sequence of cDNA

The nucleotide sequence of cDNA was analyzed by using DNA automatic sequencer 373A produced by Applied

was determined for about 750 clones which were randomly selected.

(2) Homology search

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Partial sequences of about 750 clones were searched with a computer using BlastX. As a result, three clones appeared to be homologues of bacterial cellulose synthase subunit. Therefore, it was tried to isolate full length clones.

<5> Isolation of full length clones

#### (1) 5'-RACE

As a result of the homology search, the obtained homologue clones were found to be partial length clones. Therefore, primers were synthesized to make elongation toward the 5' upstream so that RT-PCR was performed by using mRNA as a template.

#### (1-a) Synthesis of first-strand DNA

The following solution was prepared and mixed in a tube.

0.5  $\mu$ l 10  $\mu$ mol gene-specific primer 1; 1 pg total RNA; DEPC-treated H<sub>2</sub>O (adjusted to give a total amount of 9  $\mu$ l).

The following oligonucleotides were used as the gene-specific primer, 1. That is, an oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 15 was used for PcsA1. An oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 16 was used for PcsA2. An oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 17 was used for PcsA3

The reaction solution was gently mixed, and then it was centrifuged under a mild condition to allow the reaction solution to fall to the bottom of the tube. The solution was left to stand at 70 °C for 10 minutes, followed by immediate cooling on ice.

Next, the following solution was prepared and mixed in the tube.

5 x RT Buffer 5 p1; 25 mM MgCl<sub>2</sub> 2.5  $\mu$ l; 2 mM dNTP mix 5  $\mu$ l; 0.1 M DTT 2.5  $\mu$ l; H<sub>2</sub>O (added to give a total amount of 24  $\mu$ l).

The solution was gently agitated, and then it was centrifuged under a mild condition to allow the reaction solution to fall to the bottom of the tube, followed by being left to stand at 42 °C for 1 minute. The solution was added with 1  $\mu$ l of SuperScriptll RT (reverse transcriptase, GIBCO BRL), and it was gently mixed. After that, the reaction was performed at 42 °C for 50 minutes. Subsequently, the reaction solution was left to stand at 70 °C for 15 minutes to stop the reaction. Centrifugation was performed under a mild condition to allow the reaction solution to fall to the bottom of the tube, followed by being left to stand at 37 °C. RNase H (produced by Toyobo) in an amount of 1  $\mu$ l was added thereto to perform a reaction at 37 °C for 30 minutes.

Subsequently, in order to remove excessive primers and nucleotides contained in the reaction solution, gel filtration was performed by using a purification column produced by Boehringer, Quick Spin Columns. At first, the tip of the column was removed, followed by centrifugation at 1,100 x g for 2 minutes to discard the buffer. The reaction solution was introduced into the central area of the column, followed by centrifugation at 1,100 x g for 4 minutes to recover the solution.

#### (1-b) Poly(dC) tailing

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An aliquot (5 µl) was dispensed from the obtained solution, to which the following solution was added.

5 μl 5 x CoCl<sub>2</sub> Buffer:

2.5 µl 2 mM dCTP;

H<sub>2</sub>O (adjusted to give a total amount of 24 μl).

The reaction solution was mixed well, and it was left to stand at 94 °C for 3 minutes. Centrifugation was performed under a mild condition to allow the reaction solution to fall to the bottom of the tube, followed by being left to stand on ice. Terminal transferase TdT (produced by Toyobo) was added thereto in an amount of 1  $\mu$ l, followed by mixing under a mild condition to perform a reaction at 37 °C for 10 minutes. Subsequently, the reaction solution was left to stand at 65 °C for 10 minutes to stop the reaction.

#### (1-c) PCR reaction

An aliquot (2.5 µI) was dispensed from the reaction solution, to which the following solution was added.

2.5 µl 10 x PCR Buffer;

2.5 µl 2 mM dNTP mix;

0.5 μl Gene-specific primer 2;

0.5 µl Abridged Anchor Primer (GIBCO BRL);

0.5 µl Advantage Klentag Polymerase Mix (Clontech);

 $H_2O$  (adjusted to give a total amount of 25  $\mu$ I).

The following oligonucleotides were used as Gene-specific primer 2. That is, an oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 18 was used for PcsA1. An oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 19 was used for PcsA2. An oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 20 was used for PcsA3.

The solution was introduced into a 0.2 ml tube to perform the PCR reaction under the following condition.

PAD	94 °C	90 seconds
30 cycles	94 °C	30 seconds
	60 to 68 °C	30 to 60 seconds
}	68 °C	180 seconds
Final	68 °C	7 minutes
Hold	4 °C	

The reaction solution was subjected to agarose gel electrophoresis to extract, from the gel, DNA's corresponding to portions having the largest size (about 1.8 K for PcsA1, about 2 K for PcsA2, and about 2.2 K for PcsA3). GENO-BIND produced by CLONTECH was used for the extraction, and the procedure was carried out in accordance with its protocol. The DNA thus obtained was subjected to Poly(dC)tailing, which was used as a template to perform the PCR reaction. The condition and the composition of the reaction solution were the same as those described above.

### (2) Cloning

### (2-a) 5'-RACE TA cloning

Starting from the obtained PCR reaction solution, cloning was performed by using TA Cloning Kit produced by Invitrogen in accordance with its protocol.

The following solution was added to an aliquot (1.5 µl) of the PCR reaction solution obtained as described above.

0.5 μl 10 x Ligation Buffer; 1 μl pCRII vector; 0.5 μl T4 DNA Ligase; 1.5 μl dH<sub>2</sub>O.

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The reaction was performed at 14 °C overnight. An aliquot (2 p1) of the reaction solution was added to 25 p1 of Escherichia coli competent cell (JM109) preparation, followed by being left to stand for 30 minutes on ice. After that, heat shock was applied at 42 °C for 30 seconds. The solution was stationarily left to stand on ice for 2 minutes, to which 450 µl of SOB medium was thereafter added to perform cultivation at 37 °C for 1 hour with shaking at 200 rpm. The culture was spread over Amp/Xgat/IPTG plate, followed by incubation at 37 °C overnight. The plasmid was extracted from obtained colonies in accordance with the method as described above.

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The procedure was carried out by using DNA Sequencer 377 produced by ABI in accordance with its protocol. The sequencing reaction was performed by using M13 primer and synthetic oligomer as primers, based on the use of Dye Terminater Cycle Sequencing Kit produced by the same company. As a result of the sequencing, as for PcsA3, it was revealed that another clone also belonging to the group of PcsA3 but having a slightly different sequence (one position for amino acid) was isolated (see Figs. 3 and 4). A nucleotide sequence of a clone (PcsA3-682) containing the 3'-side region of PcsA3 and an amino acid sequence deduced from this nucleotide sequence are shown in SEQ ID NOs: 5 and 6. A nucleotide sequence of a 5'-portion (PcsA3-5') of another clone containing the 5'-side region of PcsA3 and an amino acid sequence deduced from this nucleotide sequence are shown in SEQ ID NOs: 7 and 8. A nucleotide sequence of a 3'-portion (PcsA3-3') of the clone and an amino acid sequence deduced from this nucleotide sequence are shown in SEQ ID NOs: 9 and 10.

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As for PcsAl and PcsA2, primers for 5'-terminal and 3'-terminal of a region containing ORF were synthesized on the basis of the obtained sequences to perform the PCR reaction. Thus, complete length clones were isolated by means of TA cloning. The condition and the composition of the reaction solution were the same as those described above.

for PcsAl. Oligonucleotides shown in SEQ ID NO: 23 (5'-terminal) and SEQ ID NO: 24 (3'-terminal) were used as the

primers for PcsA2. Results are shown in SEQ ID NOs: 1 to 4.

Oligonucleotides shown in SEQ ID NO: 21 (5'-terminal) and SEQ ID NO: 22 (3'-terminal) were used as the primers

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# Ann x to the description

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# SEQUENCE LISTING

	(1) GENERAL INFORMATION:
	(i) APPLICANT: NISSHINBO INDUSTRIES, INC.
	HAYASHI, Takahisa
10	(11) TITLE OF INVENTION: CELLULOSE SYNTHASE GENE
	(iii) NUMBER OF SEQUENCES: 24
	(iv) Correspondence address:
	(A) ADDRESSEE:
15	(B) STREET:
	(C) CITY:
	(E) COUNTRY:
	(F) ZIP:
20	(v) COMPUTER READABLE FORM:
	(A) MEDIUM TYPE: Floppy disk
	(B) COMPUTER: IBM PC compatible
	(C) OPERATING SYSTEM: PC-DOS/MS-DOS
25	(D) SOFTWARE: PatentIn Release #1.0, Version #1.30
	(vi) CURRENT APPLICATION DATA:
	(A) APPLICATION NUMBER:
30	(B) FILING DATE:
	(C) CLASSIFICATION:
	(vii) PRIOR APPLICATION DATA:
	(A) APPLICATION NUMBER: JP 9-83133
35	(B) FILING DATE: 1-APR-1997
	(viii) ATTORNEY/AGENT INFORMATION:
	(A) NAME:
	(B) REGISTRATION NUMBER:
40	(ix) TELECOMMUNICATION INFORMATION:
	(A) TELEPHONE:
	(B) TELEFAX:
	(2) INFORMATION FOR SEQ ID NO: 1:
45	(1) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 3207 base pairs
	(B) TYPE: nucleic acid
50	(C) STRANDEDNESS: double
	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: cDNA to mRNA
	(vi) ORIGINAL SOURCE:
55	(VI) ORIGINAL SOURCE:  (A) ORGANISM: Gossypium hirsutum L.
	(W) OKOWINTON' GOSSÁFTAN LITTRACTAN D'

(C) INDIVIDUAL ISOLATE: Coker312

	(ix) FEATURE:																
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5		(	B) L	OCAT:	ION:	773	3001										
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35	Tyr	Gly	Tyr	Gly	Pro	Pro	Ser	Met	Pro	Ser	Phe	Pro	Lys	Ser	Ser	Ser	•	
	540					545					550					555		
	TCA	TCT	TGC	TCG	TGT	TGC	TGC	$\infty$	GGC	AAG	AAG	GAA	CCT	AAA	GAT	CCA	1789	
	Ser	Ser	Cys	Ser	Cys	Cys	Cys	Pro	Gly	Lys	Lys	Glu	Pro	Lys	Asp	Pro		
40					560	)				56	5				57	0		
	TCA	GAG	CTT	TAT	AGG	GAT	GCA	AAA	œ	GAA	GAA	CTT	GAT	GCT	$\alpha$	ATC	1837	
	Ser	Glu	Leu	Tyr	Arg	Asp	Ala	Lys	Arg	Glu	Glu	Leu	Asp	Ala	Ala	Ile		
				575					580	כ				58	5			
45	TTT	AAC	CTT	AGG	GAA	ATT	GAC	AAT	TAT	GAT	GAG	TAT	GAA	AGA	TCA	ATG	1885	
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					640					645					65		
0					ACT												2077
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				655					660					66			
					GAT												2125
15	Ser	Val	Thr	Glu	Asp	Ile	Leu	Thr	Gly	Phe	Lys	Met	His	Cys	Arg	Gly	
5			670					675					68				
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	Trp	Arg	Ser	Ile	Tyr	Cys	Met	Pro	Leu	Arg	Pro	Ala	Phe	Lys	Gly	Ser	
		685					690					699	_		0		
20					CIG												2221
	Ala	Pro	Ile	Asn	Leu			Arg	Leu	His			Leu	Arg	Trp		
	700					705					710					715	2260
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					720					725				~~~	73		2217
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<del>1</del> 5	CAC	TTD A	m~~	CALL.	AAC		CAG	dalah	TYY			ىنچى	ഷ	بلملت			2557
					Asn												2007
	ASP	reu	urp	815		GIU	9111	FIRE	820		***	ULY.	OT.	82		- 4-1-1-1	
	CAT	CIIIC	(Intel)	-	GTC	محدمك	C A A	ىرون	_		AAC	ATY2	ملعلف			ATT	2605
50					Val												
	nus	لماستخد		and	AGT	C 1 55	ATT I	GTA	1155	u					1		

	830 835 8 <u>40</u>	
	GAC ACC AAC TIT ACT GTC ACT GCC AAA GCA GCT GAT GAT GCA GAT TIT	2653
_	Asp Thr Asn Phe Thr Val Thr Ala Lys Ala Ala Asp Asp Ala Asp Phe	
5	<b>845</b> 850 855	
	GGT GAG CTC TAC ATT GTG AAA TGG ACT ACA CTT CTA ATC CCT CCA ACA	2701
	Gly Glu Leu Tyr Ile Val Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr	
	860 865 870 875	
10	ACA CTC CTC ATC GTC AAC ATG GTT GGT GTC GTT GOC GGA TTC TOC GAT	2749
	Thr Leu Leu Ile Val Asn Met Val Gly Val Val Ala Gly Phe Ser Asp	
	GCC CIC AAC AAA GGG TAC GAA GCT TGG GGA CCA CIC TIT GGC AAA GIG	2797
15	Ala Leu Asn Lys Gly Tyr Glu Ala Trp Gly Pro Leu Phe Gly Lys Val	
	895 900 905	
	THE THE TOO THE TOO GIVE AT CITE CAT CITE TAT OCA THE CITE AAA GOT	28 <b>4</b> 5
	Phe Phe Ser Phe Trp Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly	
20	910 915 920	
	CIT ATG GGA COC CAA AAC AGG ACA CCA ACC ATT GIT GTC CTT TGG TCA	2893
	Leu Met Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Val Leu Trp Ser	
	925 930 935	
25	GTG TTG GCT TCT GTC TCT CTT GTT TGG GTT CGG ATC AAC CCG	2941
	Val Leu Leu Ala Ser Val Phe Ser Leu Val Trp Val Arg Ile Asn Pro	
	940 945 950 955	
	TIT GIC AGC AGC GCC GAT AGC AGC AGC GTG TCA CAG AGC TGC ATT TCC	2989
30	Phe Val Ser Thr Ala Asp Ser Thr Thr Val Ser Gln Ser Cys Ile Ser	
	960 965 970	3038
	ATT GAT TGT TGATGATATT ATGTGTTTCT TAGAATTGAA ATCATTGCAA	3036
	Ile Asp Cys GTAAGIGGAC TGAAACATGI CIATIGACIA AGITTIGAAC AGITTGIACC CATTITATIC	3098
35	TTAGCAGTGT GTAATTTTOC TAAACAATGC TATGAACTAT ACATATTTCA TTGATATTTA	3158
	CATTAAATGA AACTACATCA GTCTGCAGAA AAAAAAAAA AAAAAAAAA	3207
	Childhaidh ancimealch dicidenann annannanna annannan	3207
	(2) INFORMATION FOR SEQ ID NO: 2:	
40	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 974 amino acids	
	(B) TYPE: amino acid	
	(D) TOPOLOGY: linear	
45	(ii) MOLECULE TYPE: protein	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:	
	Met Met Glu Ser Gly Val Pro Val Cys His Thr Cys Gly Glu His Val	
	1 5 10 15	
50	Gly Lou Aon Val Aon Gly Gly Pro Dhe Val Ala Cre Mie Gly Cre Aon	

						20						2	:5					30	
	P	he	Pro	) Il	e C	ys 1	jys	Sea	r Cy	s Pl	he	Glu	Туз	r As	p Le	u Ly	s Gl		y Gln
5				3	35						40						45		
	L	ys	Ala	Cy )	s La	eu /	\r <del>g</del>	Cys		.y II 55	le	Pro	Туз	. As		u As 60	n Le	u Le	u Asp
	As	SP	Val	Gl	u Ly	rs A	Ца	Thr	Gl	y As	sp (	Gln	Sez	Th			a Al	a Hi	s Leu
10	. 6	55						7	0					•	75				80
	Se	r	Lys	Se	r Gl	n P	sp	Val	. G1	y Il	e i	His	Ala	Arg	J H1:	s Il	e Se	r Se	r Val
	<b>a</b> -			_	_		85						_	0					95
15	Se	ır	Thr	Lei			er	Glu	Me	t Th	ur (			) Ast	Gl <sub>3</sub>	/ As	n Pro	o Ile	e Trp
	Tay	ne:	Acn	<b>3</b> ~~		00 1 C	<b>1</b>	C	<b></b>	_	_	105					1	10	
	LJ	<b>.</b>	vəri	11!	y va 5	ı G	τu	ser	117			Glu	Lys	Lys	S Ast			s Lys	s Lys
	Pr	. מ	Ala			r I	ve	V=1	G1:		20 ~ (	~1	81 <sub>-</sub>	<b>01</b>			25		ı Gln
20			130				, 0	VUL	13		y c	<b>31</b> U	Ala	GIU	116		Pro	GII	i Gin
<b>V</b>	GL	n l	Met	Glu	ı As	рĿ	ys	Pro			o A	lsp	Ala	Ser			Len	Sar	Thr
	14	5					_	150			_			15			, 150	· Set	160
25	Ile	е :	Ile	Pro	Il	e P	က	Lys	Ser	Ar	g L	eu	Ala			Axc	Thr	· Val	Ile
						1	65						170	)				17	75
	Ile	e 1	<b>let</b>	Arg	Le	ıI	le	Ile	Leu	Gly	y L	eu	Phe	Phe	His	Тут	Arg	Val	Thr
20					18	0						185					19	0	
30	AST	) F	w	Val	Asį	) S∈	er i	Ala	Phe			eu	Trp	Leu	Thr	Ser	Val	Ile	Cys
	Gli	, T	130	195		. , , ,	_ 1	D1	<b></b>	20		. <b>.</b> .	_			20	5		
	010	• •	210	ıιρ	FIR	; AJ	.a. i	PIRE	Ser 219		) V	aı .	Leu	Asp			Pro	Lys	Trp
35	Tyr			Val	Asr	. Ar	YT (	31,,			. т	20.	N ~~~	N	220	)	Ala	_	_
	225	;					9 '	230	***	-7-	. 4.	16 1	nsp	235		ser	ATA	Arg	
	Glu	A	rg	Glu	Gly	Gl			Asn	Glu	Le	eu A	Ala			Agn	Phe	Pho	240 Val
40						24	<b>l</b> 5						250			. ພ	11.2	25	
	Ser	T	hr '	Val	Asp	Pr	o I	eu	Lys	Glu	Pr	ro I			Ile	Thr	Ala	Asn	Thr
					260	)					2	265					270	)	
	Val	L	eu S	Ser	Ile	Le	u A	la :	Leu	Asp	Ту	T F	, ox	Val	Asp	Lys	Val	Ser	Cys
45	m	<b>.</b>		275	_	_	_	_	_	280						285	5		
	TYL	1.	60 16 2	er	Asp	Ası	G	ly A	Ala	Ala	Me	et L	eu!	Ihr	Phe	Glu	Ser	Leu	Val
	Glu			la	<b>3</b> ~~	Dha		J_ ,	295		_			_	300				
50	305			ua.	vəħ	FIR	; A.	10 A	иg	гÃ2	11	pν	al I		Phe ·	Cys	Lys	Lys	
		Ιl	e G	lu :	Pm	Arr			, M	Glu	Dh	_ m	T	315	·	<b>0</b> 3	Lys		320
				•		32	, 5	I	-0	JLU	FIJ		330 330	-ue (	ser (	IΙ	ràs		
	Tyr	Le	u L	ys i	Asp			al G	ln	Pro	Sea	r Pi	be V	'al I	ive (	3111	Arg /	335	) Ala
<i>55</i>					3 <b>4</b> 0						34		•	•	, — `	- <b></b>	350		

	Met	Lys	Arg 355	Asp	Tyr	Glu	Glu	<b>Tyr</b> 360		Ile	Arg	Ile	Asn 36		Leu	Val
s	Ala	Lys 370	Ala	Gln	Lys	Thr	Pro 375		Glu	Gly	dıb	Thr 380		Gln	Asp	Gly
	Thr 385	Pro	Trp	Pro	Gly	<b>As</b> n 390		Pro	Arg	Asp	His 395		Gly	Met	Ile	Gln 400
10	Val	Phe	Leu	Gly	Tyr 405		Gly	Ala	His	Asp 410		Glu	Gly	Asn	Glu 41	
	-10	2	-Servin	11-1-	<b>M</b>	1101	Goo	Acres	-A1	Torre	-105	70-00	61-		Gla.	Udo
									-326	,						
15	His	Lys	Lys 435	Ala	Gly	Ala	Glu	Asn 440		Leu	Val	Arg	Val 44		Ala	Val
	Leu	Thr 450	Asn	Ala	Pro	Phe	Ile 455		Asn	Leu	Asp	Cys 460		His	Tyr	Val
20	Asn 465	Asn	Ser	Lys	Ala	Val 470	-	Glu	Ala	Met	Cys 475		Leu	Met	Asp	Pro 480
•		Val	Gly	Arg	Asp 485		Cys	Tyr	Val	Gln 490		Pro	Gln	Arg	Phe 49	_
25	Glv	Ile	Aso	Ara	Ser	Asp	Arq	Tyr	Ala	Asn	Arq	Asn	Thr	Val	Phe	Phe
				500		•		•	505					510		
	Asp	Val	Asn 515	Met	Lys	Gly	Leu	<b>Asp</b> 520		Ile	Gln	Gly	Pro 525		Tyr	Val
30	Gly	Thr	Gly	Cys	Val	Phe	Asn	Arg	Gln	Ala	Leu	Tyr	Gly	Tyr	Gly	Pro
	_	530	_	_			535	_				540	)			
	Pro	Ser	Met	Pro	Ser	Phe	Pro	Lys	Ser	Ser	Ser	Ser	Ser	Cys	Ser	Cys
35	545					550					555	5				560
	Cys	Cys	Pro	Gly	Lys 565	-	Glu	Pro	Lys	_	Pro	Ser	Glu	Leu	Tyr 57	
	Asp	Ala	Lys	Arg	Glu	Glu	Leu	Asp	Ala	Ala	Ile	Phe	Asn	Leu	Arg	Glu
40				580					585	5				590	כ	
	Ile	Asp	Asn 595	Tyr	Asp	Glu	Tyr	Glu 600		Ser	Met	Leu	Ile 605		Gln	Thr
	Ser	Phe	Glu	Lys	Thr	Phe	Gly	Leu	Ser	Ser	Val	Phe	Ile	Glu	Ser	Thr
45		610					615					620	)			
	Leu	Met	Glu	Asn	Gly	Gly	Val	Ala	Glu	Ser	Ala	Asn	Pro	Ser	Thr	Leu
	625					630					635	5				640
50	Ile	Lys	Glu	Ala	Ile	His	Val	Ile	Gly	Cys	Gly	Tyr	Glu	Glu	Lys	Thr
50					645					650					65	
	Ala	Trp	Gly	-	Glu	Ile	Gly	Trp		_	Gly	Ser	Val			Asp
				660					665					670	7	
	_ =	Leu				_						_	_			_

	675 680 685
	Cys Met Pro Leu Arg Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu
5	690 695 700
5	Ser Asp Arg Leu His Gln Val Leu Arg Trp Ala Leu Gly Ser Val Glu
	705 710 716
	720
	Ile Phe Leu Ser Arg His Cys Pro Leu Trp Tyr Gly Phe Gly Gly
10	725 730 735
	Arg Leu Lys Trp Leu Gln Arg Leu Ala Tyr Ile Asn Thr Ile Val Tyr
	/ <b>4</b> 0 745 750
	Pro Phe Thr Ser Leu Pro Leu Ile Ala Tyr Cys Ser Leu Pro Ala Ile
15	755 760 765
	Cys Leu Leu Thr Gly Lys Phe Ile Ile Pro Thr Leu Ser Asn Leu Ala
	770 775 780
	Ser Val Leu Phe Leu Gly Leu Phe Leu Ser Ile Ile Val Thr Ala Val
* 20	700 700 700
	Leu Glu Leu Arg Trp Ser Gly Val Ser Ile Glu Asp Leu Trp Arg Asn 805 810
25	Glu Gln Phe Trp Val Ile Gly Gly Val Ser Ala His Leu Phe Ala Val
	820 825 830
	Phe Gln Gly Phe Leu Lys Met Leu Ala Gly Ile Asp Thr Asn Phe Thr
	840 845
30	Val Thr Ala Lys Ala Ala Asp Asp Ala Asp Phe Gly Glu Leu Tyr Ile
	855 860
	Val Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Leu Leu Ile Val
	900 970
<i>35</i>	Asn Met Val Gly Val Val Ala Gly Phe Ser Asp Ala Leu Asn Lys Gly
	885 000
	Tyr Glu Ala Trp Gly Pro Leu Phe Gly Lys Val Phe Phe Ser Phe Trp
40	• 200
	Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly Leu Met Gly Arg Gln
	920 925
	Asn Arg Thr Pro Thr Ile Val Val Leu Trp Ser Val Leu Leu Ala Ser
45	935 940
45	Val Phe Ser Leu Val Trp Val Arg Ile Asn Pro Phe Val Ser Thr Ala
	74J (5)
	Asp Ser Thr Thr Val Ser Gln Ser Cys Ile Ser Ile Asp Cys
50	965 970
	(2) INFORMATION FOR SEQ ID NO: 3:
	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 3311 base point

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	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: double
-	(D) TOPOLOGY: linear
5	(11) MOLECULE TYPE: cDNA to mRNA
	(vi) ORIGINAL SOURCE:
	(A) ORGANISM: Gossypium hirsutum L.
	(C) INDIVIDUAL ISOLATE: Coker312
10	(ix) FEATURE:
	(A) NAME/KEY: CDS

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3: CTTTOGTTCT TTTGGTTTTG CC ATG GCT TCA ACC ACC ATG GCC GCT GGC TTT Met Ala Ser Thr Thr Met Ala Ala Gly Phe GGT TCA CIT GCT GTT GAC GAG AAT OGG GGA TCA TOG ACA CAT CAA TCA Gly Ser Leu Ala Val Asp Glu Asn Arg Gly Ser Ser Thr His Gln Ser TCA ACG AAA ATA TOC AGG GTG TGT GGG GAT AAG ATC GGG CAA AAG GAA Ser Thr Lys Ile Cys Arg Val Cys Gly Asp Lys Ile Gly Gln Lys Glu AAC GGA CAA COG TTC GTG GCT TGT CAT GTC TGT GCT TTC COG GTT TGC Asn Gly Gln Pro Phe Val Ala Cys His Val Cys Ala Phe Pro Val Cys CGT CCT TGT TAT GAA TAT GAA AGG AGT GAA GGA AAC CAG TGC TGT CCT Arg Pro Cys Tyr Glu Tyr Glu Arg Ser Glu Gly Asn Gln Cys Cys Pro CAG TOC AAT ACT COC TAT AAG COT CAC AAA GOT AGT CCA AGA ATT TCA Gln Cys Asn Thr Arg Tyr Lys Arg His Lys Gly Ser Pro Arg Ile Ser GGA GAT GAA GAT GAT TCA GAT CAA GAT GAT TTT GAT GAT GAA TTT Gly Asp Glu Glu Asp Asp Ser Asp Gln Asp Asp Phe Asp Asp Glu Phe CAG ATT AAG AAC COC AAG GAT GAC TOC CAT CCA CAA CAT GAA AAT GAG Gln Ile Lys Asn Arg Lys Asp Asp Ser His Pro Gln His Glu Asn Glu GAA TAT AAT AAT AAT CAT CAA TGG CAT CCC AAT GGT CAA GCT TTC Glu Tyr Asn Asn Asn Asn His Gln Trp His Pro Asn Gly Gln Ala Phe TCA GTT GOC GGA AGC AGG GOG GGG AAG GAT TTG GAA GOG GAT AAA GAG Ser Val Ala Gly Ser Thr Ala Gly Lys Asp Leu Glu Gly Asp Lys Glu

	ATT	TAC	GGA	AGC	GAA	GAA	TGG	AAA	GAA	AGA	GIT	GAG	AAA	TGG	AAA	GIC	532
	Ile	Tyr	Gly	Ser	Glu	Glu	Trp	Lys	Glu	Arg	Val	Glu	Lys	Trp	Lys	Val	
	155	_	_			160					165	5				170	
5	AGG	CAA	GAA	AAA	AGA	CCT	TTG	GTA	AGC	AAC	GAT	AAT	GGC	GGA	AAT	GAT	580
	Arg	Gln	Glu	Lys	Arg	Gly	Leu	Val	Ser	Asn	Asp	Asn	Gly	Gly	Asn	Asp	
					175					180	)				18	5	
10															CCT		628
10	Pro	Pro	Glu	Glu	Asp	Asp	Tyr	Leu	Leu	Ala	Glu	Ala	Arg	Gln	Pro	Leu	
				190					195					20			
															œ		676
15	Trp	Arg	Lys	Val	Pro	Ile	Ser	Ser	Ser	Leu	Ile	Ser			Arg	Ile	
, 0			205					210					21				
															TTC		724
	Val	Ile	Val	Leu	Arg	Phe	Phe	Ile	Leu	Ala	Phe	Phe	Leu	Arg	Phe	Arg	
20		220					225					230					620
															TCT		772
			Thr	Pro	Ala			Ala	Tyr	Pro		_	Leu	116	Ser		
	235					240					245		~~ ==	a. a	<b>~~~</b>	250	920
2 <b>5</b>															TTC		820
	Ile	Cys	Glu	Val			ALA	Pne	ser			Leu	Asp	GIN	Phe		
					255			<b>03.3</b>		260		~~~	~~	CIIIC	26		868
															TCC		800
30	Lys	Trp	Phe			'l' <b>nr</b>	Arg	GIU		_	Leu	Asp	ALG	28	Ser ∩	neu	
				270		~~	C) C	~~	275		CTOOL S	~~	~~			CTC	916
															GAC		710
	arg	Pne		_	GIU	GTĀ	GIU	290		GIII	Leu	GIY	29		ysb	VOL	
35	mmc	cmc	285		CALIFOR	CNC	بلغلب			CAA	$\sim$	arc.			ACC	CCC	964
															Thr		
	PIE	300	SEI	ш	VQI	vah	305		LIY S	GIU	110	310	1				
	አልሮ		Calair	СТА	TYY:	ልጥና			GIY	CAT	TAC			GAG	AAA	GTG	1012
40															Lys		
	315	ALG	Var	Deu		320					325					330	
		יובאני	тат	GIG	TCG			GGT	GCT	TCC			CTT	TTC	GAT	TCG	1060
															Asp		
45 .	0,0	0,0	-1-	-	335					340	_				34		
	TTG	TCT	GAA	ACG			TTC	GCG	AGG	AGA	TGG	GIT	$\infty$	TTT	TGT	AAG	1108
															Cys		
50				350					355			,		36		-	
50	AAG	CAT	AAT			$\infty$	AGG	GCG			J-J-T	TAT	TTC	AAT	GAG	AAG	1156
															Glu		

				365					370	)				37	5				
	A	TT	GAT	TAT	TTG	AAG	GAC	AAG	GTC	CAT	œr	AGC	TTT	GTT	AAA	GAA	œ	1204	
	I	le	Asp	Tyr	Leu	Lys	Asp	Lys	Val	His	Pro	Ser	Phe	Val	Lys	Glu	Arg		
5			380	-			_	385	,				390	0					
	A	GA	œ	ATG	AAA	AGG	GAA	TAT	GAA	GAA	TTT	AAA	GTA	AGG	ATC	AAT	GCA	1252	
	A	rg	Ala	Met	Lys	Arg	Glu	Tyr	Glu	Glu	Phe	Lys	Val	Arg	Ile	Asn	Ala		
	3	95					400	)				405	5				410		
10	T	TA	GTA	GCA	AAA	GCT	CAG	AAG	AAA	CCA	GAA	GAA	GGA	TGG	GIG	ATG	CAA	1300	
	L	eu	Val	Ala	Lys	Ala	Gln	Lys	Lys	Pro	Glu	Glu	Gly	Trp	Val	Met	Gln		
																	- Miles Control		
45		200	-				coc										1110		
15	A	sp	Gly	Thr	Pro	Trp	Pro	Gly	Asn	Asn	Thr	Arg	Asp	His	Pro	Gly	Met		
					430					435					44	_			
		_	_													GGC		1396	
20	I	le	Gln		Tyr	Leu	Gly	Ser			Ala	Leu	Asp			Gly	Lys		
20				445					450					45					
																GGT		1444	
	G	lu		Pro	Arg	Leu	Val			Ser	Arg	Glu		_	Pro	Gly	Tyr		
25	_		460					465				~~	470	_		~	mam.	1.400	
25																GIT		1492	
			HIS	HIS	гàг	rys		-	ATS	GIU	ASN		_	var	Arg	Val			
		75	~~~	~~~	. ~	3.30	480			3.m3	mma.	485		C) T	mæm	C M III	490	1540	
20																GAT		1540	
30	Α.	та	vai	Lesu	1111	495		PLO	FIE	TIE	500		Leu	vab	Суз	Asp 50	_		
	σr.	)AC	איזער	244	ልልሞ			arr.	ΔTY	»GC			эта	TCC	بلعلمك	TTA		1588	
												_				Leu		2000	
35	<del>-</del> ,	2-			510		-10			515				-1-	520				
00	G	ΑT	ССТ	CAG		GGA	AAG	AAG	CIT	TGT	TAT	GIT	CAA	TTT	CCA	CAG	AGA	1636	
					-											Gln			
		•		525		_	_	-	530	_	_			535					
40	T	TT	GAT	CCT	TTA	GAT	<b>CGT</b>	CAT	GAT	CGA	TAT	GCT	AAT	CGA	AAT	GTT	GIC	1684	
,,	P	he	Asp	Gly	Ile	Asp	Arg	His	Asp	Arg	Tyr	Ala	Asn	Arg	Asn	Val	Val		
			540					545					550	)					
	T	TC	TTT	GAT	ATC	AAC	ATG	TIG	GGA	TTA	GAT	GGA	CTT	CAA	œc	$\infty$ r	GTA	1732	
45	P	he	Phe	Asp	Ile	Asn	Met	Leu	Gly	Leu	Asp	Gly	Leu	Gln	Gly	$\mathbf{Pro}$	Val		
	5	55					560					565	i				570		
	T	ΑT	GTA	CCC	ACA	GCG	TGT	GIT	TTC	AAC	AGG	CAG	GCA	TTG	TAT	GGC	TAC	1780	
	T	yr '	Val	Gly	Thr	Gly	Cys	Val	Phe	Asn	Arg	Gln	Ala	Leu	Tyr	Gly	Tyr		
50						575					580	)				58	5		
	G	AT ·	CCA	CCA	GIC	TCT	GAG	AAA	CGA	CCA	AAG	ATG	ACA	TGT	GAT	TGC	TGG	1828	

	Asp	Pro	Pro	Val	Ser	Glu	Lys	Arg	Pro	Lys	Met	Thr	Cys	Asp	Cys	Trp	
	590			590					595	5				60	0		
5	CCT	TCT	TGG	TGT	TGC	TGT	TGT	TGC	GGA	GGT	TCT	AGG	AAG	AAA	TCA	AAG	1876
	Pro	Ser	Trp	Cys	Cys	Сув	Cys	Cys	Gly	Gly	Ser	Arg	Lys	Lys	Ser	Lys	
			605					610					61				
															GGA		1924
10	Lys	Lys	Gly	Glu	Lys	Lys	Gly	Leu	Leu	Gly	Gly	Leu	Leu	Tyr	Gly	Lys	
		620					625					630					
															GCA		1972
	Lys	Lys	Lys	Met	Met	Gly	Lys	Asn	Tyr	Val	Lys	Lys	Gly	Ser	Ala		
15	635					640					645					650	
															GAA		2020
	Val	Phe	Asp	Leu	Glu	Glu	Ile	Glu	Glu			Glu	Gly	Tyr	Glu		
					655					660					66		2242
20															CGA		2068
	Leu	Glu	rys	Ser	Thr	Leu	Met	Ser	Gln	rys	Asn	Phe	Glu		Arg	Phe	
				670					675					68			2116
															CCT		2116
25	Gly	Gln	Ser	Pro	Val	Phe	Ile			Thr	Leu	Met			Gly	GLY	
			685					690					69			010	2164
															ATT		2164
	Leu		Glu	Gly	Thr	Asn			Ser	Leu	Ile		_	Ala	Ile	His	
30		700					705					710			~~	3.000	2212
															GAG		2212
		Ile	Ser	Cys	Gly			Glu	Lys	Thr			GTA	rys	Glu		
	715					720					725			~~	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	730	2260
35															TTC		2260
	Gly	Trp	Ile	Tyr			Val	Thr	GIU			Leu	THE	GIY	Phe	_	
					735					740		~TD	~~		74		2308
															AGA		2306
40	Met	His	Cys			Trp	Lys	Ser			Cys	vaı	PIO		Arg	PIO	
				750		~~	~~~	. ~~	755		m	C>m	~~	76		CAA	2356
															CAC		2330
	Ala	Phe		GIĀ	Ser	ATA	PIO			Leu	Ser	veb	77!		His	0111	
45			765		~~	~~~	~~	770		CNA	N CTOTT	UAIX.			ىلتىک	CAC	2404
															OGT A		2404
	Val		Arg	Trp	ATS	Leu			var	GIU	ше			Ser	Arg	ura	
		780					785		~~=		~	790	_	Casac.	CXC	NOC.	2452
50															GAG		2432
		Pro	Leu	Trp	TYL			GTĀ	GTÅ	rÃ2			rrp	Ded	Glu		
	795					800					805	,				810	

	CIT	CCT	TAT	ATC	AAC	ACC	ATT	GIT	TAC	CCT	TTC	ACC	TOG	ATC	$\alpha$	TTA	2500
	Leu	Ala	Tyr	Ile	Asn	Thr	Ile	Val	Tyr	Pro	Phe	Thr	Ser	Ile	Pro	Leu	
5			-		815				_	820					82		
3	CTC	œ	TAT	TGT	ACT	ATT	CCA	GCT	GTT	TGT	CTT	CTC	ACC	GGC	AAA	TTC	25 <b>4</b> 8
													Thr				
			•	830					839	_				84	_		
	ATC	ATT	CCA			AGC	AAC	CIT	ACA	AGT	GTG	TGG	TTC	TTG	GCA	CTT	2596
10													Phe		_		
P			845					850					85				
_	Phe	Leu	Ser	Ile	Ile	Ala	Thr	Glv	Val	Leu	Glu	Leu	Arg	Tro	Ser	Gly	
15		860	-				865					870	_				
	بلملك	-	ATC	CAA	CAC	TYCG			ААТ	GAA	CAA		TGG	GTG	ATC	GGA	2692
													Trp				
	875	-	116	011.	rup.	880		9			885			•		890	
20		CITC	TVC-	~	ChT			CCT	CITY.	محلمك			CIC	CITC	222		2740
																	2740
	GTĀ	vai	Ser	Ala His Leu Phe 895				ALG.	Vall	900		GLY	Deu	LEU	90		
	CTT 3	~~	CC3	CTUA			220	יראוארי	λCC			CCA	AAA	CC N	-		2788
25																	2766
	Leu	АТа	GTĀ		_	The	ASI	Prie			THE	ALA	Lys			veh	
	~~ ~		~~~	910		~~	~~~	ma m	915			m~~	202	92		OWN	2026
													ACA				2836
30	Asp	Inr		Pne	GIĀ	GLU	Leu			Pne	rys	ııp	Thr		Leu	Leu	
			925					930					93			~~~	2004
													GGA				2884
	Ile		Pro	Thr	Thr	Leu			Leu	Asn	Met		Gly	Val	Val	Ala	
35		940					945					950					2000
													TGG				2932
	_	Val	Ser	Asp	Ala			Asn	Gly	Tyr		,	Trp	Gly	Pro		
	955					960					965					970	
40													CAT				2980
	Phe	Gly	Lys	Leu	Phe	Phe	Ala	Phe	_			Leu	His	Leu			
					975					980	)				98	5	
	TTC	CTC	AAA	GGT	TIG	ATG	GGG	AGA	CAA	AAC	AGG	ACG	$\infty$	ACC	ATT	GTT	3028
45	Phe	Leu	Lys	Gly	Leu	Met	Gly	Arg	Gln	Asn	Arg	Thr	Pro			Val	
43				990					995	i				100	00		
	GTG	CTT	TGG	TCC	ATA	CIT	TIG	GCA	TCG	ATT	TTC	TCA	CIG	GTT	TGG	GTA	3076
	Val	Leu	Trp	Ser	Ile	Leu	Leu	Ala	Ser	Ile	Phe	Ser	Leu	Val	Trp	Val	
50			1005	5				101	0				101	15			
50	œ	ATC	GAT	$\infty$	TTC	TTG	$\infty$	AAA	CAA	ACA	<b>GGT</b>	CCA	GIT	CTT	AAA	CAA	3124
	Ara	Ile	Asp	Pro	Phe	Leu	Pro	Lys	Gln	Thr	Gly	Pro	Val	Leu	Lys	Gln	

	1020 1025 1030													
	TGT GGC GTG GAG TGC TAAATGGTGT TTTACAAACC TTTCTTATTA TTTTATTTTC	3179												
5	Cys Gly Val Glu Cys													
	1035													
	OCTIVITION ACTIVITIES ATTRICTOR ATTRICTOR													
	CCTTTTGCC ACTACTGTTG ATTTGCTGTG ATTCTAAAAG GGATTTATCT TGTTTGTAAA													
	AAGTCTCCTA TGATTTTGTT GGTTCAATTT AATTTCTATA TGGTAAAAAA ATATTTCTTT AAATTAACTA TA													
10	ANNI TANCTA TA	3311												
	(2) They may no one of													
	(2) INFORMATION FOR SEQ ID NO: 4:													
	(1) SEQUENCE CHARACTERISTICS:													
15	(A) LENGTH: 1039 amino acids													
	(B) TYPE: amino acid													
	(D) TOPOLOGY: linear													
	(ii) MOLECULE TYPE: protein													
20	(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 4:													
20	Met Ala Ser Thr Thr Met Ala Ala Gly Phe Gly Ser Leu Ala Val Asp													
	5 10													
	Glu Asn Arg Gly Ser Ser Thr His Gln Ser Ser Thr Lys Ile Cys Arg													
	20													
25	23 30													
	Val Cys Gly Asp Lys Ile Gly Gln Lys Glu Asn Gly Gln Pro Phe Val													
	<del></del> 0 45													
	Ala Cys His Val Cys Ala Phe Pro Val Cys Arg Pro Cys Tyr Glu Tyr 50 55													
30	50													
	Glu Arg Ser Glu Gly Asn Gln Cys Cys Pro Gln Cys Asn Thr Arg Tyr													
	75 80													
	Lys Arg His Lys Gly Ser Pro Arg Ile Ser Gly Asp Glu Glu Asp Asp													
35	85 90 95													
	Ser Asp Gln Asp Asp Phe Asp Asp Glu Phe Gln Ile Lys Asn Arg Lys													
	100 105 110													
	Asp Asp Ser His Pro Gln His Glu Asn Glu Glu Tyr Asn Asn Asn													
	115 120 125													
40	His Gln Trp His Pro Asn Gly Gln Ala Phe Ser Val Ala Gly Ser Thr													
	135 140													
	Ala Gly Lys Asp Leu Glu Gly Asp Lys Glu Ile Tyr Gly Ser Glu Glu													
	190 190 190 190 190 190 190 190 190 190													
45	Trp Lys Glu Arg Val Glu Lys Trp Lys Val Arg Gln Glu Lys Arg Gly													
	100													
	4/0 1/5													
	Leu Val Ser Asn Asp Asn Gly Gly Asn Asp Pro Pro Glu Glu Asp Asp													
50	100													
	Tyr Leu Leu Ala Glu Ala Arg Gln Pro Leu Trp Arg Lys Val Pro Ile													
	195 200 205													

		Ser	Ser	Ser	Leu	Ile	Ser	Pro 215		Arg	Ile	Val	11e 220		Leu	Arg	Phe	
			210	_			_,	_		<b>5</b> 1		<b>71</b> _			D-00	N1 a	Tr	
٤	5	Phe 225	Ile	Leu	Ala	Phe	230		Arg	Pne	Arg	235		TITE	PIO	Ata	240	
			Ala	Tyr	Pro	Leu	Trp	Leu	Ile	Ser	Val	Ile	Cys	Glu	Val	Trp	Phe	
				•		245					250		_			25		
1	o	Ala	Phe	Ser	Trp	Ile	Leu	Asp	Gln	Phe	Pro	Lys	Trp	Phe	Pro	Ile	Thr	
	_				260					265					27			
		Arg	Glu	Thr	Tyr	Leu	Asp	Arg	Leu	Ser	Leu	Arg	Phe	Glu	Arg	Glu	Gly	
1	5	Glu	Pro	Asn	Gln	Leu	Gly	Pro	Val	Asp	Val	Phe	Val	Ser	Thr	Val	Asp	
			290				_	295		_			300					
		Leu	Leu	Lvs	Glu	Pro	Pro	Ile	Ile	Thr	Ala	Asn	Ala	Val	Leu	Ser	Ile	
		305					310					315					320	
2	0		Ala	Va1	Asn	ጥረም			Glu	Lvs	Val			Tvr	Val	Ser		
		Deu	Au	V	, mp	325			0_0	-1-	330		-1-	-1-		33		
		۸	Gly	λla	Sor			Lau	Pha	Aen			Ser	Glu	Thr			
		ASŲ	Gry	ALG	340	MEC	LEU	Deu		345				014	35		<b>0-2</b>	
2	₽5	T16	11-	3		<b>T</b>	Ual	D	Dha			Tare	Wie.	Aen			Pro	
		Pne	Ala	_	Arg	пр	van	PIO			пуз	пуз	ms	36		OLU	110	
		_		355		<b></b> 1	<b></b>	<b>70</b> L -	360		T	<b>71</b> ~	*			T + 200	Acn	
		Arg	Ala	Pro	GIU	Pne	ıyr			GIU	rys	me			Leu	БУЗ	voh	
3	30		370		_			375			_	_	380		T	3	Cl.	
		_	Val	His	Pro	Ser			ràs	GIU	Arg			Met	гÃ2	ALY		
		385					390			_		395				•••	400	
		Tyr	Glu	Glu	Phe			Arg	Ile	Asn			Val	Ата	ràs			
3	35					405					410					41		
		Lys	Lys	Pro	Glu	Glu	Gly	Trp	Val	Met	Gln	Asp	Gly	Thr	Pro	Trp	Pro	
					420					425		1			43			
		Gly	Asn	Asn	Thr	Arg	Asp	His	Pro	Gly	Met	Ile	Gln	Val	Tyr	Leu	Gly	
4	10			435					440	)				44	5			
		Ser	Ala	Gly	Ala	Leu	Asp	Val	Asp	Gly	Lys	Glu	Leu	Pro	Arg	Leu	Val	
			450					455	,				460	)				
		Tyr	Val	Ser	Arg	Glu	Lys	Arg	Pro	Gly	Tyr	${\tt Gln}$	His	His	Lys	Lys	Ala	
4	<b>1</b> 5	465					470					475	5				480	
		Gly	Ala	Glu	Asn	Ala	Leu	Val	Arg	Val	Ser	Ala	Val	Leu	Thr	Asn	Ala	
		_				485			_		490	)				<b>4</b> 9	5	
		Pro	Phe	Ile	Leu	Asn	Leu	Asp	Cys	Asp	His	Tyr	Ile	Asn	Asn	Ser	Lys	
5	50				500			•	_	505		-			51			
		Ala	Met	Ara		Ala	Met	Cys	Phe			Asp	Pro	Gln			Lys	
				515		- <b></b>		-2-	520			- 2-		52		-	<del>-</del>	
		[ws	Leu		Tvr	Val	Gln	Phe			Ara	Phe	Asp			Asp	Arg	
	5 <i>5</i>	273	عانب	J, U	-1-						3		P	1		- &-	•	